

AMERICAN ASSOCIATION OF VETERINARY LABORATORY DIAGNOSTICIANS EVENT

68th Annual Meeting, October 30-November 5, 2025, Aurora, Colorado.

AAVLD AWARDS

E.P. Pope Award

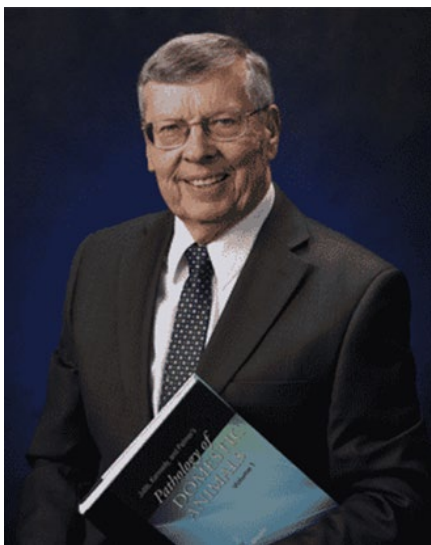


Dr. Amy K. Swinford, DVM, MS, Diplomate (Emeritus) ACVM

Amy K. Swinford is director of the Texas A&M Veterinary Medical Diagnostic Laboratory. Dr. Swinford is a four-time graduate of the University of Illinois, earning bachelor's degrees in both microbiology and veterinary science, a DVM, and a master's degree in veterinary pathobiology. She became a diplomate of the American College of Veterinary Microbiologists in 1997. She started her career as a diagnostic virologist at the University of Nebraska Veterinary Diagnostic Center in Lincoln, studying pseudorabies virus and what was then a recently discovered disease, porcine reproductive and respiratory syndrome (PRRS). During this time, Dr. Swinford also served as a public health officer in the United States Air Force Reserve.

Read more... [Biography](#)

Distinguished Service Award



M. Grant Maxie, DVM, Dip Sm An Med, PhD, Diplomate ACVP

Grant Maxie was born and raised in southern Alberta. After graduating with his DVM from the Western College of Veterinary Medicine in Saskatoon in 1969, he completed an internship in small animal medicine and a PhD in clinical pathology at the Ontario Veterinary College in 1973. Following 3 years working in Kenya on a bovine trypanosomosis (sleeping sickness) research project, he joined the faculty at the OVC as an anatomic pathologist, teaching cardiovascular and urinary pathology.

Read more... [Biography](#)

Outstanding Performance Award for Diagnostic Services



Mary Payton, BHS, MBA MT(ASCP)

Mary Payton has been at UKVDL since 2005. First, as a Clinical Pathology Supervisor. Then, in 2008, QA Manager

Previously, I had my own physician office and hospital consulting service, which I created Procedure Manuals, QA programs, Safety Policy, and provided training for CLIA, JCAHO, COLA, CAP, and Ky Safety OSHA. I was also a sales representative for Infolab, a small laboratory and healthcare product company.

From 1980-1992, I was the Microbiology and Serology Supervisor at St. Joseph Hospital in Lexington, Ky.

Read more... [Biography](#)

AAVLD President's Award



Alexis Thompson, DVM, Ph.D., DACVPM (Epidemiology)

Alexis Thompson is the Resident Director of the Texas A&M Veterinary Medical Diagnostic Laboratory (TVMDL) in Canyon, TX. She first joined TVMDL in 2023 as a bovine veterinary diagnostician and was promoted to resident director in 2024. In 2015, Thompson earned a bachelor's degree in animal science from the University of Kentucky. She earned a Doctor of Veterinary Medicine degree in 2019, completed a residency in Population and Preventive Medicine in 2022, and earned her doctorate in 2023, all from Mississippi State University. Thompson is boarded in veterinary preventive medicine (2023) and epidemiology (2024).

Read more... [Biography](#)

Life Membership Award



CLAIRE BUCHANAN ANDREASEN, DVM, PhD, Diplomate ACVP

Read more... [Biography](#)



Dr. William G. Van Alstine, DVM, PhD,

Dr. Van Alstine received his DVM from the University of Missouri, College of Veterinary Medicine in 1981. After working in a predominantly large animal practice in Minnesota, he attended Iowa State University and received a PhD in anatomic pathology in 1987. He was board-certified by the American College of Veterinary Pathologists and joined the faculty of Purdue University College of Veterinary Medicine and Indiana Animal Disease Diagnostic Laboratory that same year.

Read more... [Biography](#)



Robert H. Poppenga, DVM, PhD.

Robert H. Poppenga received my DVM and PhD degrees from the University of Illinois in 1978 and 1987, respectively. During my PhD studies, I was also a staff veterinarian for the National Animal Poison Control Center at the College of Veterinary Medicine (the center subsequently became the ASPCA Animal Poison Control Center). Since receiving my PhD and residency training in veterinary toxicology, I have held faculty positions at Michigan State University, University of Pennsylvania, and University of California at Davis (UCD) veterinary schools. I am currently a Full Professor, Emeritus, having been a veterinary toxicologist at the California Animal Health and Food Safety Laboratory System (CAHFS), School of Veterinary Medicine, UCD, from 2004 to 2025. I retired from CAHFS on July 1, 2025.

Read more... [Biography](#)



François Elvinger, Dr.med.vet., Ph.D., dipl. ACVPM & ECVPH

François Elvinger, Professor of Veterinary Epidemiology, Associate Dean for Diagnostic Operations and Government Relations, and Executive Director of the Cornell University College of Veterinary Medicine Animal Health Diagnostic Center and New York State Veterinary Diagnostic Laboratory, obtained his veterinary and Dr.med.vet. degrees at the Hannover Veterinary School in Germany (1981 and 1983), his Ph.D. from the University of Florida (1990), before taking his first faculty position as veterinary epidemiologist at the University of Georgia Veterinary Diagnostic and Investigational Laboratory in Tifton, Georgia (1990-1997), then moving on to the Large Animal Clinical Sciences Department and Production Management Medicine group at the Virginia-Maryland College of Veterinary Medicine at Virginia Tech in Blacksburg, Virginia (1997-2015).

Read more... [Biography](#)



Dr. Mo Salman, DVM, MPVM, PhD, DACVPM, FACE, Hon. Dipl. AVOHS,

Dr. Mo Salman is a distinguished professor in the Department of Clinical Sciences at Colorado State University (CSU) and the founder and director of the Animal Population Health Institute within the College of Veterinary Medicine and Biomedical Sciences. With over 40 years of experience, Dr. Salman has an extensive educational background in veterinary medicine (BVMS), preventive veterinary medicine (MPVM), and comparative pathology (PhD). He is a Diplomate of the American College of Veterinary Preventive Medicine (ACVPM), a Fellow of the American College of Epidemiology (ACE), and an Honorary Diplomate of the American Veterinary One Health Society (AVOHS).

Read more... [Biography](#)



Dr. William Wilson, Ph.D.,

Dr. Wilson obtained his BSc (honors, 1979) and PhD (1985) in Animal Science at the University of Illinois. He was a Cromwell Postdoctoral Fellow before joining the USDA, ARS, Arthropod-Borne Animal Diseases Research Laboratory (ABADRL), located in Laramie, WY, as a Research Microbiologist in 1986. Dr. Wilson was relocated to Manhattan, KS, in 2020 with the Arthropod-Borne Animal Diseases Research Unit (ABADRU). He transferred to the Foreign Arthropod-Borne Animal Diseases Research Unit (FABADRU) at the National Bio and Agro-defense Facility (NBAF) in 2020 before retiring in December 2024.

Read more... [Biography](#)



Dr. Amy K. Swinford, DVM, MS, Diplomate (Emeritus) ACVM

Amy K. Swinford is director of the Texas A&M Veterinary Medical Diagnostic Laboratory. Dr. Swinford is a four-time graduate of the University of Illinois, earning bachelor's degrees in both microbiology and veterinary science, a DVM, and a master's degree in veterinary pathobiology. She became a diplomate of the American College of Veterinary Microbiologists in 1997. She started her career as a diagnostic virologist at the University of Nebraska Veterinary Diagnostic Center in Lincoln, studying pseudorabies virus and what was then a recently discovered disease, porcine reproductive and respiratory syndrome (PRRS). During this time, Dr. Swinford also served as a public health officer in the United States Air Force Reserve.

Read more... [Biography](#)

JVDI Best Full Scientific Report



Michael S. Filigenzi

Mike Filigenzi, BA, served as the manager of the Toxicology Laboratory at the California Animal Health and Food Safety Laboratory when he retired in 2023. Mike graduated with a Bachelor's Degree in Psychobiology from Pitzer College and shortly thereafter began working in the then-nascent contract environmental laboratory industry.

Presentation:

"Mass spectrometry in animal health laboratories: recent history, current applications, and future directions"

Read more... [Biography](#)

JVDI Best Brief Report



Savannah R. Charnas, DVM, MPH

Savannah holds a Doctor of Veterinary Medicine (DVM) degree from the University of Florida College of Veterinary Medicine and a Master of Public Health (MPH) degree from the University of Florida School of Public Health & Health Professions. She is currently serving as a third-year veterinary toxicology resident at the Kansas Veterinary Diagnostic Laboratory, where her responsibilities include interpreting diagnostic cases and educating third and fourth-year veterinary students.

Presentation:

"Acute outbreak of polioencephalomalacia in adult beef cows on pasture associated with feeding broiler litter"

Read more... [Biography](#)

Scientific Session Oral Presentation Award-1st Place



Brooklyn Cauwels

My name is Brooklyn Cauwels, and I am a Diagnostic Associate II in the Serology section of the Iowa State University Veterinary Diagnostic Laboratory. I received a BS in Biology, minor in Chemistry from ISU, and have completed my second year of graduate school at ISU, pursuing a MS in Immunobiology.

Presentation:

"Detection of Influenza A virus NP antibodies in bovine milk: evaluation of sample processing and assay repeatability using a commercial blocking ELISA"

Read more... [Biography](#)

Scientific Session Oral Presentation Award-2nd Place



Chloe Stenkamp-Strahm, DVM, PhD.

Chloe Stenkamp-Strahm has a DVM and PhD from Colorado State University, and a research background studying ruminant herd health and infectious diseases. Her post-doctoral work at Colorado State University focused on measuring impacts of the highly pathogenic avian influenza H5N1 virus in U.S. dairy cows, including studying patterns of herd and individual animal level detection of virus over time.

Presentation:

"Dairy H5N1: Insights from an early detection study of herds in California."

Read More... [Biography](#)

Scientific Session Best Poster Award -1st Place



Bala Mounika Reddi

Bala Mounika Reddi is currently a graduate student in Dr. Rahul K. Nelli's laboratory at Iowa State University. Her research centers on advancing molecular diagnostic workflows in veterinary diagnostic laboratories, with an emphasis on high-throughput RT-qPCR platforms and automation for large-scale detection of animal pathogens.

Presentation:

"Automated 384-well RT-rtPCR assay for high-volume diagnostic testing of porcine reproductive and respiratory syndrome virus (PRRSV)"

Read more... [Biography](#)

Scientific Session Best Poster Award-2nd Place



Sree Varsha Kuduva Sankar Babu (Varsha Sankar)

Sree Varsha is a graduate student pursuing a master's degree in Comparative Biomedical Sciences at the College of Veterinary Medicine, Oklahoma State University. She holds a degree in Veterinary Medicine and a Postgraduate Diploma in Veterinary Laboratory Diagnosis, with professional experience in veterinary clinical pathology.

Presentation:

"Bacteriophages as potential alternatives to treat multidrug-resistant *Pseudomonas aeruginosa* infections"

Read more... [Biography](#)

Pioneer in Virology Award



Dr. William Wilson, Ph.D.,

Dr. Wilson obtained his BSc (honors, 1979) and PhD (1985) in Animal Science at the University of Illinois. He was a Cromwell Postdoctoral Fellow before joining the USDA, ARS, Arthropod-Borne Animal Diseases Research Laboratory (ABADRL), located in Laramie, WY, as a Research Microbiologist in 1986. Dr. Wilson was relocated to Manhattan, KS, in 2020 with the Arthropod-Borne Animal Diseases Research Unit (ABADRU). He transferred to the Foreign Arthropod-Borne Animal Diseases Research Unit (FABADRU) at the National Bio and Agro-defense Facility (NBAF) in 2020 before retiring in December 2024.

Read more... [Biography](#)

Award for Excellence in Diagnostic Veterinary Microbiology Supported by BIOMIC



Josh Daniels, DVM, PhD, DACVM, Bacteriology/Myology)

Josh Daniels is a diagnostic microbiologist and has served as section head of the Bacteriology/Myology section of The Colorado State University Veterinary Diagnostic Laboratory since 2017. Prior to his time at Colorado State, Josh was the head of Clinical Microbiology at The Ohio State University Veterinary Teaching Hospital from 2008-2017. Josh trained at the University of Wisconsin and Washington State University for his DVM and PhD, respectively. He has authored or contributed to over 75 published manuscripts and book chapters.

Read more... [Biography](#)

Award for Excellence in Technical Veterinary Microbiology Supported by BIOMIC



Karen Olsen, BS

Karen is a Minnesota native and a graduate of the University of Minnesota, where she earned her degree in Microbiology. She has devoted 38 years to the University of Minnesota Veterinary Diagnostic Laboratory, beginning as a technician in the bacteriology lab and working her way up to Section Manager.

Read more... [Biography](#)

Molecular Diagnostics & Bioinformatics Oral Presentation Award (Technology Committee)



Dr. Suzanna Storms, DVM, PhD

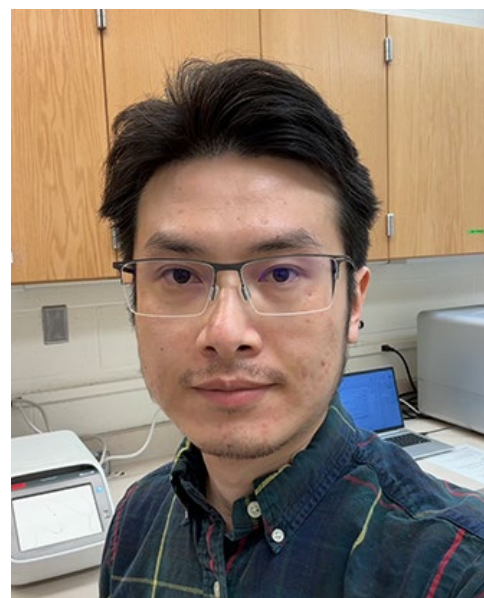
Dr. Suzanna Storms, DVM, PhD, is a Clinical Microbiology Resident at the University of Illinois Veterinary Diagnostic Laboratory. Her professional focus includes diagnostic assay development and infectious disease surveillance in livestock populations.

Presentation:

“Case report of highly pathogenic avian influenza H5N1 detected in a cat and rats during a backyard poultry outbreak.”

Read more... [Biography](#)

Molecular Diagnostics & Bioinformatics Poster Presentation Award (Technology Committee)



Hao Tong

Hoa received his bachelor's degree in veterinary medicine from Huazhong Agricultural University in China in 2015. From 2015 to 2018, he worked at his family farm as a veterinarian. In 2018, he began pursuing an M.S. in Veterinary Preventive Medicine at the VDPAM department at Iowa State University.

Presentation:

“Comparative analysis of PRRSV shedding patterns in weaned pigs vaccinated with six MLV strains: A multi-sample assessment under experimental conditions.”

Read more... [Biography](#)

J. Lindsay Oaks award, bacteriology, best student molecular biology presentation-poster



Mostafa Shelkamy, PhD

Mostafa is a PhD candidate in Veterinary Microbiology and Preventive Medicine at Iowa State University, working with the ISU poultry team. His research focuses on developing and improving molecular diagnostic tools for avian pathogens, with an emphasis on *Avibacterium paragallinarum*, the causative agent of infectious coryza.

Presentation:

“Genomic characterization and virulence assessment of non-pathogenic *Avibacterium paragallinarum* isolates from healthy layer chickens”

Read more... [Biography](#)

Walker Award for best oral bacteriology presentation



Ana Fiorella Castillo-Espinoza

Ana earned her DVM from the National University of San Marcos, Peru, and an MSc in Veterinary Preventive Medicine at Iowa State University (ISU), where she studied *Mycoplasma hyopneumoniae*-airway interactions using a three-dimensional (3D) tracheal model.

Presentation:

“Organotypic 3D tracheal model reduces time to viability detection of *Mycoplasma hyopneumoniae* compared to bacterial culture: A pilot study using tracheal swabs of known status.”

Read more... [Biography](#)

Love Award for best bacteriology poster



Nneka Vivian Iduu

Nneka is a doctoral researcher in the Biomedical Sciences Program at the College of Veterinary Medicine, Auburn University, with a focused interest in molecular diagnostics, antimicrobial resistance, and infectious diseases.

Presentation:

“High-resolution melting curve FRET-qPCR rapidly distinguishes *Streptococcus equi* subsp. *equi* and *zooepidemicus*.”

Read more... [Biography](#)

Dr. David Bemis Microbiology Trainee Travel Award



Diba Yaghoubi

Diba Yaghoubi is a graduate student in Dr. Viju Pillai's lab in the Department of Comparative Pathobiology at Purdue University College of Veterinary Medicine. I earned my bachelor's degree in cellular and molecular biology from Iran, followed by a master's degree in integrative physiopathology from École Pratique des Hautes Études – PSL Research University in Paris, France.

Presentation:

Three outbreaks of cocklebur (*Xanthium* spp.) plant poisoning in goats in Northern California

Read more... [Biography](#)

Pathology slide presentation awards First Place



Calvin Ko

Calvin is a mycoplasmologist and a veterinary pathologist dedicated to swine disease research and diagnostics. His specialties include bacterial culture and modern diagnostic tools application for fastidious Mollicutes, and he is now a Graduate Assistant Researcher and Anatomic Pathology Resident at Iowa State University.

Presentation:

"Porcine astrovirus type 4 tracheitis and bronchitis."

Read more... [Biography](#)

Pathology slide presentation awarded Second Place



Naomi Falconnier

Naomi is an anatomic veterinary pathologist with a special interest in invertebrate pathology and molecular diagnostics. She studies infectious diseases of decapod crustaceans at the University of Florida as graduate student.

Presentation:

"White spot syndrome virus in a cohort of imported blue crayfish (*Procambarus alleni*)"

Read more... [Biography](#)

Pathology slide presentation awards Third Place



Trevor Arunsiripate

Trevor Arunsiripate is a veterinarian in my third year of a combined anatomic pathology residency and PhD at Iowa State University's Veterinary Diagnostic Laboratory. All of my education has come from ISU. I am from undergrad, veterinary school, and now residency, and a graduate degree.

Presentation:

“Glomerulopathy in a 2-year-old Dachshund.”

Read more... [Biography](#)

AAVLD/ACVP Resident Diagnostic Pathology Travel Award



Margaret Odom

Margaret grew up in Seattle, Washington, and earned my DVM at Cornell University. Prior to entering the field of pathology, she completed a small animal rotating internship in Seattle, and she is now a second-year anatomic pathology resident at the Texas A&M Veterinary Medical Diagnostic Laboratory.

Presentation:

“Aberrant migration of *Angiostrongylus Cantonensis* in the central nervous system of a capybara.”

Read more... [Biography](#)

ACVP/AAVLD Resident Diagnostic Pathology Travel Award



Naomi Falconnier

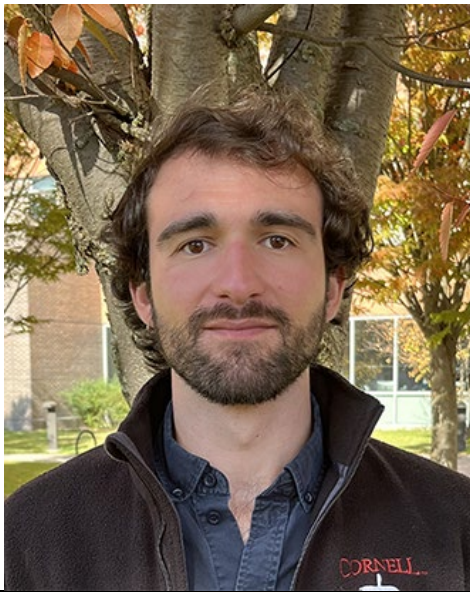
Naomi is an anatomic veterinary pathologist with a special interest in invertebrate pathology and molecular diagnostics. She studies infectious diseases of decapod crustaceans at the University of Florida as a graduate student.

Presentation:

“White spot syndrome virus in a cohort of imported blue crayfish (*Procambarus alleni*)”

Read more... [biography](#)

Pathology Oral Presentation Award



Dr. Agustin Rebollada-Merino

Agustin is a veterinarian born and raised in an agricultural region of southwestern Spain, where the central role of animals in society inspired him to pursue a degree in Veterinary Medicine (DVM) and later a Ph.D.

Presentation:

“Pathology of listeriosis (*L. monocytogenes*) in cats: a case series.”

Read more... [Biography](#)

AAVLD Laboratory Staff Travel Awards



Amy McKinney

Amy is a lab manager at the Athens Veterinary Diagnostic Laboratory at the University of Georgia, where she has worked for the past 17 years. She began her career as a bench technician in bacteriology, where she developed a passion for veterinary diagnostics and quality laboratory practices.

Presentation:

“Validation of a Next Generation Sequencing workflow for identification of fungal pathogens and antifungal resistance.”

Read more... [Biography](#)

Trainee Travel Awardees



Jingyi (Jenny) Li

Jingyi is a second-year anatomic pathology resident at the University of Illinois Urbana-Champaign. She earned her DVM from Tufts University Cummings School of Veterinary Medicine in 2024 and is originally from Shanghai, China. Her professional interests include diagnostic pathology, infectious and zoonotic diseases, and digital pathology.

Presentation:

“Histopathological Characterization of a Diarrheic Boer Goat Kid Case and Identification of a Novel Caprine Astrovirus With mNGS.”

Read more... [Biography](#)



Dr. Suzanna Storms, DVM, PhD

Dr. Suzanna Storms, DVM, PhD, is a Clinical Microbiology Resident at the University of Illinois Veterinary Diagnostic Laboratory. Her professional focus includes diagnostic assay development and infectious disease surveillance in livestock populations.

Presentation:

“Case report of highly pathogenic avian influenza H5N1 detected in a cat and rats during a backyard poultry outbreak.”

Read more... [Biography](#)



Daniela Hidalgo-Montealegre, DVM

Daniela Hidalgo-Montealegre, DVM, is a second-year master’s student in Comparative Biomedical Sciences at The Ohio State University College of Veterinary Medicine. Her research focuses on antimicrobial resistance and the surveillance of multidrug-resistant pathogens, particularly Carbapenem-Resistant Enterobacterales.

Presentation:

“Colonization with carbapenem-resistant Enterobacterales: Prevalence and risk factors in patients admitted to a companion animal intensive care unit.”

Read more... [Biography](#)



Mostafa Shelkamy, PhD

Mostafa is a PhD candidate in Veterinary Microbiology and Preventive Medicine at Iowa State University, working with the ISU poultry team. His research focuses on developing and improving molecular diagnostic tools for avian pathogens, with an emphasis on *Avibacterium paragallinarum*, the causative agent of infectious coryza.

Presentation:

“Genomic characterization and virulence assessment of non-pathogenic *Avibacterium paragallinarum* isolates from healthy layer chickens”

Read more... [Biography](#)



MaRyka Smith, PhD.

MaRyka is from the Center for Outcomes Research and Epidemiology within the College of Veterinary Medicine at Kansas State University. I am a PhD Candidate under Dr. Mike Sanderson. I completed my Doctor of Veterinary Medicine in May 2024 and hope to finish my PhD next spring.

Presentation:

“Network-based evaluation of cattle traceability with EID tags.”

Read more... [Biography](#)



Ana Fiorella Castillo-Espinoza

Ana earned her DVM from the National University of San Marcos, Peru, and an MSc in Veterinary Preventive Medicine at Iowa State University (ISU), where she studied *Mycoplasma hyopneumoniae*-airway interactions using a three-dimensional (3D) tracheal model.

Presentation:

“Organotypic 3D tracheal model reduces time to viability detection of *Mycoplasma hyopneumoniae* compared to bacterial culture: A pilot study using tracheal swabs of known status.”

Read more... [Biography](#)



Maria Chaves, PhD.

Maria is a PhD Candidate in Veterinary Microbiology at Iowa State University. She obtained her DVM in Brazil and moved to the United States in 2022 to pursue her dreams of higher education and improving her research skills. Her work focuses on the application of molecular diagnostic tools, specifically, Next Generation Sequencing Platforms for infectious agents’ identification and characterization.

Presentation:

“Nested PCR Enrichment Improves Molecular Characterization of Poultry Viruses from Clinical Samples.”

Read more... [Biography](#)

Bacteriology/Foundation sponsored



Calvin Ko

Calvin is a mycoplasmatologist and a veterinary pathologist dedicated to swine disease research and diagnostics. His specialties include bacterial culture and modern diagnostic tools application for fastidious Mollicutes, and he is now a Graduate Assistant Researcher and Anatomic Pathology Resident at Iowa State University.

Presentation:

“Development of a rapid flow cytometry-based antimicrobial susceptibility test for *Mycoplasma hyopneumoniae*.”

Read more... [Biography](#)

Epidemiology/Foundation sponsored



Dr. Berenice Munguía-Ramírez, DMV

Berenice obtained her DVM degree in México in 2019. She completed her MSc in Veterinary Preventive Medicine at Iowa State University in 2022, and currently, she is a PhD Candidate in Population Sciences in Animal Health at Iowa State University under the mentorship of Dr. Zimmerman and Dr. Giménez-Lirola.

Presentation:

“Active participatory surveillance for transboundary pathogens - Can we afford it?”

Read more... [Biography](#)

Pathology/Foundation sponsored



Pernille Knos Jorgensen (PJ), DVM

Pernille received her DVM from Copenhagen University in Denmark. She spent the first 5 years after graduation working in clinical practice in Norway and the United Kingdom. Following [TE1] this, she worked as a Veterinary Investigation Officer in Scotland for 6 years, performing necropsies on farm animals.

Presentation:

“Three outbreaks of cocklebur (*Xanthium* spp.) plant poisoning in goats in Northern California.”

Read more... [Biography](#)

Virology/Foundation sponsored



Konner Cool, PhD.

Konner is a PhD student in the Department of Diagnostic Medicine/Pathobiology at Kansas State University's College of Veterinary Medicine.

Presentation:

“Susceptibility of dry cattle to infection with highly pathogenic avian influenza H5N1.”

Read more... [Biography](#)

Center of Public and Corporate Veterinary Medicine (CPCVM) Awardees



Stefanie Coates

Stefanie was raised in Raleigh, North Carolina, surrounded by her parents, sister, three cats, two dogs, and two horses. She graduated from Davidson College in 2019 with a double major in Biology and German Studies.

Lincoln Memorial University College of Veterinary Medicine

Class of 2027

Read more... [Biography](#)



Sarah Harden Hiser, PhD/DVM

Sarah is a candidate at NC State's College of Veterinary Medicine, where she focuses on infectious disease dynamics and antimicrobial resistance.

NC State University College of Veterinary Medicine

Class of 2027

Read more... [Biography](#)



Michelle Lund, DVM

Michelle is a candidate in the Class of 2029 at the Virginia-Maryland College of Veterinary Medicine, as well as a Registered Veterinary Technician.

Virginia-Maryland College of Veterinary Medicine

Class of 2029

Read more... [Biography](#)



Cassie Paterson

Cassie is a current first-year veterinary student at the University of Minnesota with interests in anatomic pathology and community medicine. She earned her Bachelor of Science in Biological Sciences at the University of Pittsburgh in 2022 and is originally from the Philadelphia area.

University of Minnesota College of Veterinary Medicine

Class of 2029

Read more... [Biography](#)



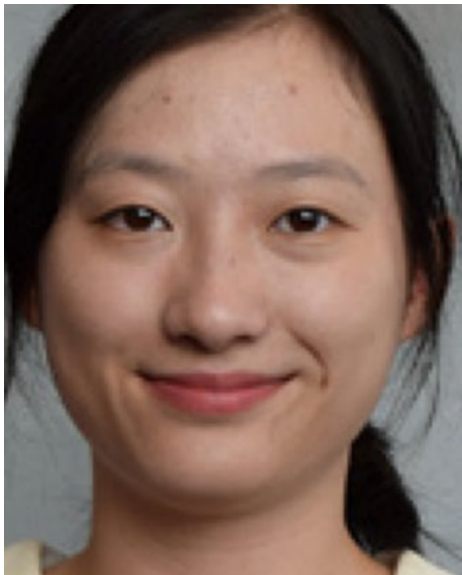
Jus Tupik

Jus is a current third-year DVM student tracking public/corporation at the Virginia-Maryland College of Veterinary Medicine. She finished her PhD at Virginia Tech in 2024 on the innate immune response to Brucellosis and Lyme Disease.

Virginia-Maryland College of Veterinary Medicine

Class of 2027

Read more... [Biography](#)



Jiamao Ysng

Jiamao is a second-year student at Iowa State University CVM. her hometown is Chongqing, a vibrant city of mountains, rivers, and bridges, situated in the southeast of China.

Iowa State University College of Veterinary Medicine

class of 2027

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OFFICIALS

Drs. Tanya LeRoith, Blacksburg, Virginia, president; Ashley Hill, Davis, California, president-elect and program chair; Francisco Uzal, San Bernardino, California, vice president; Kristy Pabilonia, Fort Collins, Colorado, secretary-treasurer; Keith Poulsen, Madison, Wisconsin, immediate past president; and David H. Zeman, Brookings, South Dakota, executive director

Amy K. Swinford, DVM, MS, Diplomate (Emeritus) ACVM, is director of the Texas A&M Veterinary Medical Diagnostic Laboratory. Dr. Swinford is a four-time graduate of the University of Illinois, earning bachelor's degrees in both microbiology and veterinary science, a DVM, and a master's degree in veterinary pathobiology. She became a diplomate of the American College of Veterinary Microbiologists in 1997. She started her career as a diagnostic virologist at the University of Nebraska Veterinary Diagnostic Center in Lincoln, studying pseudorabies virus and what was then a recently discovered disease, porcine reproductive and respiratory syndrome (PRRS). During this time, Dr. Swinford also served as a public health officer in the United States Air Force Reserve.

In 2005, Dr. Swinford joined the Texas A&M Veterinary Medical Diagnostic Laboratory (TVMDL) as section head of diagnostic bacteriology/mycology. She also enjoyed collaborative research, and serving as an adjunct assistant professor in the College of Veterinary Medicine & Biomedical Sciences at Texas A&M University, where she delivered lectures in veterinary microbiology and infectious disease to first through third-year veterinary students.

Dr. Swinford became TVMDL's microbiology branch chief in 2012, overseeing the bacteriology, virology, serology, and molecular diagnostics sections. As branch chief, she provided leadership, oversaw stringent quality assurance and safety programs, and implemented new testing protocols. She became particularly interested in the quality assurance standards that applied to veterinary diagnostic laboratories, which led her to join the AAVLD Accreditation Committee's Auditor Pool, and ultimately the AAVLD Accreditation Committee. She enjoyed many years serving on the Committee, auditing AAVLD laboratories, and seeing the AAVLD Accreditation Program Standard in action as it enhanced the quality of veterinary diagnostics across the United States.

Dr. Swinford became associate director of TVMDL in 2015 and was appointed TVMDL's sixth agency director in 2021. During her tenure, Dr. Swinford coordinated TVMDL's responses to numerous disease outbreaks such as vesicular stomatitis, contagious equine metritis, anthrax, chronic wasting disease, highly pathogenic avian influenza, and SARS-CoV-2. She helped implement TVMDL's testing of human patients for SARS-CoV-2 during the COVID-19 pandemic and guided her laboratory's response to the outbreak of highly pathogenic avian influenza in dairy cattle, first discovered in the Texas Panhandle in 2024.

Dr. Swinford has been recognized with awards from the Texas A&M University System, including the TVMDL Director's Excellence Award, three Vice Chancellor's Awards in

Excellence, and the Texas A&M System Regents Fellow Service Award. On November 13, 2025, the Texas A&M System Board of Regents bestowed the title of director emeritus in recognition of Dr. Swinford's 20-plus years of dedicated service to TVMDL and its clients.

Dr. Swinford has been an active member of AAVLD since 1992, attending her first AAVLD/USAHA Annual Meeting in 1993 in Las Vegas, Nevada. She's had the privilege of serving on numerous committees, including Accreditation, Subcommittee on Bacteriology and Mycology, Subcommittee on Antimicrobial Susceptibility Testing, Membership, Government Relations, Laboratory Directors, Strategic Planning, AAVLD/USAHA Joint Committee on the NAHLN, AAVLD Committee Chairs and Program Committee, AAVLD House of Delegates, and the AAVLD Executive Board.

Grant Maxie, DVM, PhD, Diplomate ACVP

Formerly:

Director, Animal Health Laboratory, and Co-Executive Director, Laboratory Services Division,
University of Guelph, Guelph, Ontario, Canada
(retired, August 31, 2019)

Phone: (519) 822-1640. Cell 519-835-3481

Email: gmaxie@uoguelph.ca

Grant Maxie was born and raised in southern Alberta. After graduating with his DVM from the Western College of Veterinary Medicine in Saskatoon in 1969, he completed an internship in small animal medicine and a PhD in clinical pathology at the Ontario Veterinary College in 1973. Following 3 years working in Kenya on a bovine trypanosomosis (sleeping sickness) research project, he joined the faculty at the OVC as an anatomic pathologist, teaching cardiovascular and urinary pathology. After moving to the Veterinary Services Laboratory of OMAFRA in Guelph in 1982, he became board-certified in anatomic pathology by the American College of Veterinary Pathologists in 1984. He worked as a diagnostic pathologist and then as Guelph laboratory head for VLSB until 1997. For 22 years he was the Director of the Animal Health Laboratory plus, for the last 12 years, co-Executive Director of the Laboratory Services Division at the University of Guelph and retired in 2019.

Pursuing an interest in scientific editing, Grant was the editor-in-chief of the Canadian Veterinary Journal (1986-91), has published extensively, and co-authored chapters in the 3rd to 7th editions and edited the 5th, 6th, and 7th editions of "*Jubb, Kennedy, and Palmer's Pathology of Domestic Animals*". He is a past-president of the American Association of Veterinary Laboratory Diagnosticians (AAVLD, 2007-08), volunteered for many years as an assessor on the Accreditation Committee, and is the current editor-in-chief of the *Journal of Veterinary Diagnostic Investigation* (2015-present).

Grant married Dr. Laura Smith-Maxie (veterinary neurologist, also now retired) in Kenya in 1975. They have 2 children – Kevin, entrepreneur and showjumper, Erin, ON (first grandchild, Tristan Eames, 2020; granddaughter Coco Yvaine, 2025); Andrea, teacher and outdoorsperson, Nelson, BC. Grant is enjoying retirement, keeps editing and consulting, continues farming, plus horseback trail-riding with his wife.



Mary Payton, BHS, MBA MT(ASCP) has been at UKVDL since 2005. First, as a Clinical Pathology Supervisor. then, in 2008, QA Manager

Previously, I had my own physician office and hospital consulting service, which I created Procedure Manuals, QA programs, Safety Policy, and provided training for CLIA, JCAHO, COLA, CAP, and Ky Safety OSHA. I was also a sales representative for Infolab, a small laboratory and healthcare product company.

From 1980-1992, I was the Microbiology and Serology Supervisor at St. Joseph Hospital in Lexington, Ky.

After graduating from the UK College of Allied Health as a Medical Technologist, I worked in Chemistry, Hematology, and Microbiology at two other hospitals in Lexington.

I graduated from the UK with a BHS in 1976. I received my MBA from Xavier University in 1988.

I have 3 dogs - two Rottweilers and one Jack Russell, and a very proud Gigi to 3 grandchildren. I live in Versailles, Ky, with my husband, Heath, and the three dog "children".

CURRICULUM VITAE

CLAIRE BUCHANAN ANDREASEN, DVM, PhD, Diplomate ACVP

Address: Department of Veterinary Pathology
College of Veterinary Medicine
Iowa State University
Ames, IA 50011
candrease@iastate.edu

EDUCATION AND TRAINING

1987-1990	PhD	Pathology, University of Georgia, Athens, Georgia Dissertation: <i>Heterophil function in healthy chickens and in chickens with staphylococcal tenosynovitis.</i>
1985-1987	MS	Pathology, University of Georgia, Athens, Georgia Thesis: <i>Intermediate filament staining in the histologic and cytologic diagnosis of canine skin and soft tissue tumors.</i>
1979-1982	DVM	Texas A & M University, College Station, Texas
1981	BS	Veterinary Medicine, <i>Summa Cum Laude</i> , Texas A & M University, College Station, Texas
1975-1979	BS	Biomedical Science, <i>Summa Cum Laude</i> , Texas A & M University, College Station, Texas

BOARD CERTIFICATION

1994 Diplomate, American College of Veterinary Pathologists (Clinical Pathology)

RESEARCH EMPHASIS

Comparative clinical pathology/pathology, granulocyte cell function (emphasis; poultry, fish), cellular host-pathogen interaction (emphasis: infectious disease)

EDUCATION/LEARNING/OUTREACH/PROFESSIONAL PRACTICE EMPHASIS

Clinical pathology/comparative clinical pathology, Infectious disease pathology, images for infectious diseases (emerging, foreign to U.S., zoonotic <http://www.cfsph.iastate.edu/DiseaseInfo/disease-images.php>)

ADMINISTRATIVE EMPHASIS

Departmental - Integration of research, diagnostics, and DVM and graduate education; director of pathology laboratories services.

Work-life faculty issues associated with the ISU Office of the Provost. Past responsibilities for grants, meeting presentations, committees, resources. Example: Resource Guide for Recruiting Excellent and Diverse Faculty

EMPLOYMENT/TITLE

2025-present	University Professor, Emerita, Iowa State University (ISU), Ames, Iowa
2021- 2024	University Professor, Iowa State University (ISU), Ames, Iowa
May 2022- 2025	Clinical Pathology Laboratory Director, ISU, Ames, Iowa
2018-2025	Clinical Pathology residency coordinator, Veterinary Pathology, ISU, Ames, Iowa
May 2023-Jan 2024	Director of Graduate Education (DOGE), Veterinary Pathology, ISU, Ames, Iowa
2018- 2022	Professor, Dr. Roger and Marilyn Mahr Professorship in One Health, ISU, Ames, Iowa
2016-2022	Director of One Health, ISU, Ames, Iowa

2010- 2016 Professor and Associate Dean for Academic and Student Affairs, ISU,, Ames, Iowa.

2003-2025 Professor, Clinical Pathology, Veterinary Pathology, ISU, Ames, Iowa.

2008 – 2009 Interim Chair, Department of Clinical Sciences, ISU, Ames, IA

2006 Special Assistant to the Provost, Office of the Provost, ISU, Ames, IA.

2005-2006 Administrative Intern (50%), Office of the Provost, directed by the Associate Provost for Academic Advancement and Diversity, ISU, Ames, Iowa.

2001-2011 Chair, Department of Veterinary Pathology, ISU, Ames, Iowa.

2001 Interim chair, Department of Veterinary Pathology, ISU, Ames, Iowa.

2000- 2011 Director of the Departmental Clinical Pathology Laboratory, ISU, Ames, Iowa

1999-2003 Associate Professor, Clinical Pathology, Veterinary Pathology, ISU, Ames, Iowa.

1996-1999 Assistant Professor, Clinical Pathology, Veterinary Pathology, ISU, Ames, Iowa.

1995-1996 Assistant Professor, Clinical and Surgical Pathology, Veterinary Diagnostic Laboratory and College of Veterinary Medicine, Oregon State University (OSU), Corvallis, Oregon.

1994-1996 Section Head, Clinical Pathology Laboratory, Veterinary Diagnostic Laboratory and College of Veterinary Medicine, OSU, Corvallis, Oregon.

1993-1994 Co-director, Clinical Pathology Laboratory, Veterinary Diagnostic Laboratory and College of Veterinary Medicine, OSU, Corvallis, Oregon.

1991-1994 Assistant Professor, Clinical and Anatomic Pathology, Veterinary Diagnostic Laboratory and College of Veterinary Medicine, OSU, Corvallis, Oregon.

1990-1991 Instructor, Clinical and Anatomic Pathology, Veterinary Diagnostic Laboratory and College of Veterinary Medicine, OSU, Corvallis, Oregon.

1990 Instructor, Clinical Pathology, Department of Veterinary Pathology, University of Georgia, College of Veterinary Medicine, Athens, Georgia.

1985-1990 Veterinary Medical Graduate Assistant, Clinical Pathology and Surgical Biopsy, Department of Veterinary Pathology, University of Georgia, College of Veterinary Medicine, Athens, Georgia.

1982-1985 Associate Veterinarian, Northeast Veterinary Clinic, 9405 Dyer, El Paso, Texas

HONORS AND AWARDS

Alpha Lambda Delta, 1975-1976
 Phi Kappa Phi, 1979
 Who's Who Among American College Students, 1979
 Gamma Sigma Delta, 1979
 Pre-Veterinary Society Scholarship Award, 1979
 Distinguished Student Certificates, 1975-1980
 Spring Award Scholarship, 1979-1982
 The National Dean's List, 1981
 Hill's Nutrition and Dietary Management Award, 1982

Phi Zeta, 1982-present

Membership Committee, 1994-1996

Nomination Committee 1996

SmithKline Beecham (Pfizer) Award for Research Excellence, 1994

SCAVMA Clinical Science Teaching Award, 2001

Osborn Club Scientific Organization (by invitation), Iowa State University, elected 2003/2004

ISU Foundation Outstanding Achievement in Departmental Leadership 2005

Women Impacting ISU Calendar, Iowa State University 2013

Texas A&M University College of Veterinary Medicine and Biomedical Sciences Outstanding Alumnus 2018

Dr. Roger and Marilyn Mahr Professorship in One Health 2018

Marshal, ISU President Wintersteen Installation 2018

ISU University Professor, 2021

CVM Frederick Douglass Patterson Diversity and Inclusion Award, 2021

CVM University of Georgia Distinguished Graduate Award, 2022

EDITORIAL BOARDS

Board of scientific reviewers for **American Journal of Veterinary Research** 1993-1996.

Editorial Board **Veterinary Pathology** 1997-1999.

Editorial Board **Comparative Clinical Pathology** 2002-2004; 2016-present

Review Editor in Veterinary Humanities and Social Sciences, **Frontiers in Veterinary Science**, 2016-2020.

Review Editor in Veterinary Experimental and Diagnostic Pathology, **Frontiers in Veterinary Science** 2020-present

PROFESSIONAL ORGANIZATIONS

Past:

American Animal Hospital Association, 1981-1985

Texas Veterinary Medical Association, 1982-1984

El Paso Veterinary Medical Association, 1982-1985

Continuing Education Chairman, 1982-1984

Texas Academy of Veterinary Practice, 1982-1985

American Association of Feline Practitioners, 1983-1985

Faculty Advisor Oregon State 1996

Willamette Valley Veterinary Medical Association 1991-1996

Oregon Veterinary Medical Association, 1994-1995

Member of Midwest Consortium for Toxicologic Pathology 2003-2011

Iowa Chapter of the Office of Women in Higher Education, 2005-2010

Internal Advisory Committee for Education and Outreach, National Center for Foreign Animal and Zoonotic

Disease Defense (FAZD), Texas A&M University, 2010-2015

American Association of Veterinary Medical Colleges

Chair of Constituent Group of Chairs, Heads, and Directors of the AAVMC 2004-2006

Committee for the Vision in Veterinary Medicine Foresight Analysis Project 2006

Associate Deans for Academic Affairs

Chair-elect 2012-2013

Chair 2013-2014

Past Chair 2014-2015

Committee for AAVMC Teaching Award, Committee Chair, 2014, 2015

World Aquatic Veterinary Medical Association 2007- 2014, 2016 founding member

Co-lead for Education in the Center of Excellence for Emerging and Zoonotic Animal Diseases (CEEZAD) (multi-institutions; KSU- overall lead) 2010-2016.

Advisor: Council of Advisors, Gerson Lehman Group's Council of Healthcare Advisors. 2003-2019

Current:

American Veterinary Medical Association, 1982-Present

Iowa Veterinary Medical Association 2001-present

IVMA Foundation board 2010- 2016

IVMA Public Health committee, attendee 2018-2020; member 2020-present

C. L. Davis Foundation
 Associate Member, 1985-1990
 Member, 1995-1998, 2001-2014, ISU member, 2014- present

American Society for Veterinary Clinical Pathology, 1985-Present
 Avian Hematology Committee, 1993-1994
 Development Committee, 1998-1999
 Executive Board Committee, 1996-2001
 President-elect 1999
 President – 2000
 Past President – 2001
 ASVCP Mock Exam writer and submitter for ISU 2020 - present

American Association of Veterinary Laboratory Diagnosticians, 1991-Present
 American Association of Avian Pathologists 2004-present
 American College of Veterinary Pathologists 1994-present
 Pilot Participant for Developmental Activities 1994-1999
 ACVP Training representative for ISU clinical pathology, 2001- 2011, 2018 - present
 Specialty Session co-chair for Emerging Diseases Session, 2003 ACVP meeting
 ACVP Credentialing Committee 2002-2004, Chair 2004 -2007
 QA/QC ACVP Board Certification Committee 2003-2004
 Government Relations Committee 2007
 Credentials Committee 2009-2011
 Development Committee 2014
 President Elect 2010-2011
 President 2011-2012
 Past-President 2012-2013; Officer Nominations Committee, Chair
 Past Past President 2013-2014; Distinguished and Honoree Member Committee, Chair
 Distinguished and Honoree Member Committee, Member 2014-2015
 ACVP/STP Coalition
 Strategic Planning Committee 2010-2011
 ACVP Liaison 2011-2012
 Appeals Committee, member, 2016-present

American Association of Veterinary Medical Colleges, DEI Committee, member 2020- present
 National institute of Anti-Microbial Resistance Research and Education (NIAMRRE), member 2018-present
 Agro/Biodefense Regional Consortium liaison Coalition for Epi Response, Engagement and Science (CERES) 2018-2022
 American Society of Investigative Pathology (ASIP) 2020-present
 Conference of Research Workers in Animal Diseases 2000-2022

TEACHING RESPONSIBILITIES

Iowa State University

Instructor-in-charge

2018- present	Special Topics, Clinical Pathology rounds (VPTH 590), weekly each semester
2001- 2008	Introduction to Veterinary Cytology (Vpth 409), (15 hours/year)
1999, 2001, 2004	Pathologic Hematology (Vpth 652) (45 hours/year) Graduate hematology
2000 - 2001	Clinical Pathology (Vpth 425), core course, (70 hours/year)
2000	Clinical Pathology elective (Vpth 408), (15 hours/year) clinical pathology case solving
1996-2000	Clinical Pathology (Vpth 457), cytology and hematology senior DVM students
2002-2008	1 credit, 13 times (260 hours/year) 1996-1999

	1 credit, 10 times/year (200 hours/year) 1999-2000, (100 hours/year)
1998-2000	Freshman Honors Mentoring Program, spr, (HON 290H) 3 credits total, 2 students 1998 1 credit, 1 student 1999 1 credit each, 2 students 2000 1 credit each, 2 students 2017 1 credit each, 3 students 2018
1998-2011	Special Topics (Vpth 490), 1-4 credits, on demand
1997-2011, 2018-present	Special Topics in Clinical Pathology (Vpth 590D) (30 hours/year) Resident Rounds/Seminar
1997-2000	Introduction to Clinics (VCS 440) (21 hours/year); Cytology laboratory section
Participating Instructor	
2020- 2024	Clinical Pathology (Vpth 425), leukocyte lectures (5-7)
2024	Equine Diagnostic Skills (VCS 7447), hematology lectures (2)
2008 – present	Clinical Pathology (Vpth 457), cytology, chemistry, hematology senior DVM students
1997,1999, 2000, 2018, 2021, 2022	Seminar Spring, Fall (Vpth 604) - clinical pathology cases
2017-2020	Introduction to Veterinary Cytology and Laboratory Techniques (Vpth 409). spring. 1-2 lecture, labs/year
2017-present	Systemic Pathology (VPTH 571), fall, hematopoietic lymphoid system lectures
2017-2019	Clinical Intern seminars (2-3 yearly)
Oct 2016, 2017	VDPAM 410 Llama Medicine; Llama clinical pathology (1 hour)
Nov 2011- 2015	Veterinarian in Society III VCS 313 Ethics (2 hour lecture; 1 hour 2014) Social Media
April 2004	Special Topics in Analytical Chemistry (Chem 619); Forensic science seminar series. Wildlife forensics; how to solve the case.
Nov 2003	Animal Science (211), ISU, Issues facing Animal Science: “Animal and Environmental Welfare – The Swedish Perspective” C Andreasen, Sara Hooge (VM4) and Russ Voigt (VM3)
1997, 1999, 2002	Avian Anatomy (VANat 416) - Avian hematology (2 hours/year)
1997, 2001	Advanced Medicine (VCS 676) (1 hour lecture, 2 hours lab/year)
1996, 1999, 2001	National Board Review-clinical pathology (2 hours/year)
1996	Introduction to Pet Birds (VCS 405)-clinical pathology (2 hours/year)
Oregon State University	
1996	General Pathology (VM 511/621) (4 hours/year), disturbances of circulation

1995 Clinical Diagnosis (VM 790-20) (30 hours/year), clinical pathology application

1995 Advanced Clinical and Diagnostic Veterinary Toxicology (VM 781-11) clinical pathology case section (3 hours/year)

1994-1996 Large Animal Gastroenterology (VM 790-26), clinical pathology/techniques section (3 hours/year)

1994-1996 Neonatal Medical Care (VM 790-27), clinical pathology/techniques section (3 hours/year)

1994-1996 Microscopic Anatomy (VM 714)), hematology section (5 hours/year)

1993-1996 Camelid Medicine and Surgery (VM681), clinical pathology section (2 hours/year)

1995-1996 Clinical Services Block 1--Clinical Pathology (VM 736),(160 hours/year)

1994-1995 Clinical Services Block 1--Clinical Pathology (VM 736), (240 hours/year)

1991 -1994 Clinical Services Block 1--Clinical Pathology (VM 736),(160 hours/year); Orientation for Clinical Services Block 1 (Clinical pathology and necropsy techniques) (8 hours/year)

1993-1996 Advanced Clinical Pathology (VM 781B), senior veterinary students (40 hours/year)

1990-1994 Clinical Services Block I--Necropsy (VM 736) and senior veterinary students (180 hours/year)

University of Georgia

1990 Supervision of graduate student training in pathology.

1987-1990 Clinical Pathology (VPP 540), senior veterinary students

1987-1990 Clinical Pathology Laboratory (VPP 525), sophomore veterinary students.

1986-1989 Clinical Pathology Laboratory (VPP 421L), junior veterinary students.

Selected teaching seminars attended

March 1993 Faculty Development Program, Teaching with a case-based curriculum (AAHA), 1993

October 1996 Seldin Teaching Seminar, Center for Teaching Excellence, Iowa State University

October 1996 Teaching with Style, College of Veterinary Medicine, Iowa State University

October 1997-2001 Project LEA/RN, Iowa State University, monthly class.

International Experiences- Mentoring, Education

2001 Faculty Mentor: 4 senior veterinary students attending the Small Animal World Congress, Vancouver, Canada.

2002 Site visit to Uppsala, Sweden for 2003 student trip to understand EU food animal welfare issues.

2003 Faculty Mentor: Student trip to Sweden; 14 students; Agricultural Practices in Sweden. International trip to Sweden: <http://www.vetmed.iastate.edu/academics/international/recenttrips/sweden2003/> (inactive URL)

2007 Presentation and deliver educational materials.at the First Regional Conference on Education- Balkan Education Conference, USDA Faculty Exchange Program.

2007 OIE Vaccination for the control of Avian Influenza, Verona, Italy. Organization of educational materials for High Pathogenic Avian Influenza, sponsored by USDA and ISU Center for Food Security and Public Health.

ADVISING/MENTORING

Graduate Program membership

Member Department of Veterinary Pathology Graduate Program, 1996-present
Member Immunobiology Interdepartmental Graduate Program, Iowa State University, 1998-present
Admissions Committee, 1999-2000
Supervisory Committee, 2000-2003
Member Graduate Faculty, Iowa State University, 1996-present
Member Forensics Certificate Program, Iowa State University, 2003-2017, Graduate Program - present
Member Graduate Faculty, Oregon State University, 1991-1996

Undergraduate/DVM Advising/Mentoring

Oregon State University

Veterinary Student Advisor:

1993 Sara Anne Lange - Senior paper: Masticatory muscle myositis in the canine.
1994 Sandi Statter - Senior paper: Urinary incontinence in the bitch.
1994 Sandi Statter - Hepatic Encephalopathy - 2nd Place, Hill's Nutrition Award
1994 Susan Neary Mayo - Senior Manuscript Competition - **Veterinary Medicine**: Mayo SN, Andreasen CB, Long P: Uroperitoneum and Renal Hypoplasia in a Cria.

Iowa State University

Summer Agricultural Minority Program (undergraduate). Summer 1999.
Rolanda Morris from New Mexico, Native American Reservation. Project: Studies of avian hematology on the Cell Dyne and manual hemacytometer counts and differentials. Presentation and published proceedings required.

Freshman Honors Mentoring Program (HON 290H) (undergraduate)

Individual course program focused on students who are interested in the field of veterinary medicine and biomedical sciences.

1998 – 2 students in areas of veterinary research.

1999 - correlation of automated and manual avian hematology parameters and differential leukocyte counts.
Student: Jackie Henry.

2017 – 2 students for One Health: Megan Conry – Surgery, clinical skills, and physical therapy, the human and animal connection (co-mentors Dr. Cerfogli, Ms. Joanna Hildreth); Marydith Barkley – Human impact on raptors (Wildlife Care Clinic – ISU CVM co-mentor Dr. Zaffarano) and poster.

2018 – 3 students for One Health: Grace Mercer – Fracture location, treatment, and recovery outcomes (Wildlife Care Clinic – ISU CVM, co-mentor Dr. Zaffarano); Leah Johnson – Metarubricytosis- pathogenesis in humans and canines; Zoie Green – canine and human animal bond, evaluation of BP during brief interactions (co-mentor, Dr. Millman Hartline).

1999- Work study student learning immunohistochemistry techniques; Shawna Greene.

2010- present AgDiscovery student (ages 15-17) activities summer program- Coordinate and present - Hematology laboratory- sponsored by USDA-APHIS.

2014 - 4H Summer program 5-8 grades veterinary careers, coordinate a day for careers and activities

2016-present – Veterinary Careers for GW Carver summer interns, CALS, presentation

General academic advisor to DVM students:

1996-2001 Iowa State University College of Veterinary Medicine
DVM Student advisor

Faculty mentor for student AAV Chapter, ISU 2004-2010
Faculty advisor for student United Humanitarian Council - 2017

Merck/Merial and NIH T32 Summer Research Scholars Program

2001 Co-director. Merck/Merial and NIH Summer Veterinary Research Scholars Program.

2002 Director. Merck/Merial and NIH Summer Veterinary Research Scholars Program.

Students:

1998 Leigh Rasmussen (student), C Andreasen. Role of ascorbic acid in regulating *in vitro* granulocyte (neutrophil and heterophil) bacterial killing. Results presented at the Conference for Research Workers in Animal Diseases (CRWAD).

2000 Shawna Greene (student), C Andreasen. Comparison of greyhound platelet function and platelet counts to non-greyhounds. Results presented at Conference for Research Workers in Animal Diseases (CRWAD).

2002 Tyson Marshall (student), C Andreasen, J Greenlee. Decreased platelet numbers accompanied by gross lesions and other clinicopathologic abnormalities in a guinea pig model of leptospirosis.

2004 Dawn Herolt, (student) D Palić, C Andreasen, J Roth. Toxic agent exposure and immune cell response in fat head minnows. 1st place ACVP Annual Meeting Veterinary Student Poster Competition, 2004.

2005 Rachel Tell (student), D Palić, C Andreasen, J Roth. Cell function and research modeling in zebrafish.

2006 EM. Godinez (student), D Palić, C Andreasen, J Roth. Cytochemical and functional characterization of neutrophils from bluegills.

2007 KD Vaagen (student), Andreasen C B, Roth J A, Palic, D. Immunomodulatory Effects of Yeast Cell Wall Preparations on Neutrophil Function in Fathead Minnows (*Pimephales promelas* Rafinesque, 1820).

2018 Elizabeth Hines (student), Andreasen CB. Comparative Hematology in a One Health Approach – Pathophysiology and Prognosis of Inappropriate Metarubricytosis of Humans and Canines

2020 Aletta Anderson (student), Andreasen CB. Establishing Data-based Reference Intervals and Health Status for Metarubricytosis (nRBCs) in Dogs

2020 Gabriella K Henthorn (student), Van Vertloo L, Andreasen CB. Hematologic Findings in Cats and Dogs PCR positive for *Bartonella sp.* (ISU CVM poster 3rd place, 2020 Summer Scholars)

2022 Elisabeth Amerson (student), Andreasen CB. Cytochemical staining of hamster neutrophils. (ISU CVM 4th place award, 2022 Summer Scholars)

General mentoring:

2004 ISU Preparing Future Faculty Mentoring Program (Dusan Palić)

2005 ISU Student Scholars and Leaders Recognition Ceremony (Elise Huffman)

Graduate Students:

Oregon State University (year of completion)

Graduate Student Committee, Co-major (acting) - Mohammed Anwer, 1992

MS Thesis: *Effect of treatments on heterophil chemotaxis*

Iowa State University (year of completion)

Major Professor: Denise Wunn, Veterinary Pathology MS, 2000

The interaction of Shiga toxin with erythrocytes and endothelial cells

Major Professor: Karen Kline, Veterinary Pathology MS, 2001

Differentiation of diseases that cause mononuclear pleocytosis

Co-major Professor: Justin Greenlee, Veterinary Pathology PhD, 2003

Mechanisms in cellular pathogenesis of Leptospirosis

Co-major Professor: Dusan Palić Immunobiology PhD, 2005 [acting onsite co-major professor (Dr. Menzel was in D.C.)]

*Cytochemical and functional characterization of neutrophils from kidney of fathead minnow (*Pimephales promelas* Rafinesque 1820) and potential for use as indicator of environmental health*

ISU graduate student research excellence award, spring 2004

Co-major Professor: Shannon Hostetter, Veterinary Pathology PhD, 2009

Cellular and cytokine responses in a swine model of E Coli O157:H7

NIH KO8 2006-2009

Co-major Professor: Phongsakorn Chuammitri, IMBIO, Immunobiology PhD, 2010

Effects of genetic background and dietary immunomodulators on chicken heterophil function and Salmonella resistance

Major Professor: Ariel Nenninger, Veterinary Pathology PhD, 2023. Co-major May 2022- Feb 2023, Major professor Feb 2023-May 2023. (J. Smith/M. Ellinwood laboratory) Evaluation of a canine model of MPS 1 and MPS 111B for ocular pathology

Graduate Student Program of Study (POS) Committees: (year of completion)

Zahar Radi, Veterinary Pathology PhD, 1999

The effects of neutrophil-mediated damage in acute Mannheimia (Pasteurella) haemolytica pneumonia

Amanda Fales-Williams, Veterinary Pathology PhD, 2000

Conditions that influence production of anionic peptide in ruminant epithelium

Julie Ridgway, VCS non-thesis MS, 2000 (acting major professor)

Differentiation of histiocytic and lymphocytic neoplasia

Janice Endsley, Immunobiology PhD, 2000

Evaluation of antigen specific T cell subset responses to bovine viral diarrhea virus and bovine herpes virus-1

Jessica Caverly, Veterinary Pathology PhD, 2001

The effects of pneumonia on antimicrobial production

Tim Morgan Veterinary Pathology PhD, 2003

Mechanisms of E. coli O157:H7 Shiga toxin passage through intestinal mucosa

Brett Sponseller, Molecular, Cellular, and Developmental Biology PhD, 2003

Genetic and phenotypic characterization of the equine infectious anemia virus surface unit envelope glycoprotein during disease progression

Dianna Jordan Veterinary Pathology PhD, 2003

Novel E Coli O157:H7 colonization factors

JoAnn Morrison Veterinary Clinical Sciences, MS, 2008, (acting major professor)

Cytologic correlation of mast cell neoplasia

Laura Baseler Animal Science, Meat Science, MS, 2009 *Effects of natural curing on meat quality*

Sarah Beth Genetics, Animal Science, PhD, 2010 *Genetic and dietary effects on chicken heterophil function and immune response to Salmonella enteritidis*

Redmond Immunobiology, PhD, 2012 *Zebrafish immunology, cell function, and mycobacterial interactions.*

Rachel Tell Interdepartmental Genetics, Department of Animal Science, PhD, 2018, *Chicken genetics in heat stress resistance*

Kaylee Rowland Veterinary Pathology PhD, 2025

Cheng-Shun Hsueh *Establishment and Validation of In Situ Diagnostic Assays and a Unified Genotypic Classification System for Avian Reoviruses*

Carly Kanipe Immunobiology/Veterinary Microbiology and Preventive Medicine, PhD, pending, *Interactions of BCG and Mycobacterium bovis*

International PhD students

Inger Lilliehook completed April 1999- Opponent for PhD candidate: Inger Lilliehook, College of Veterinary Medicine, University of Agriculture, Uppsala, Sweden. Dissertation: *Studies of blood eosinophils and neutrophil granulocytes in healthy and diseased dogs.*

Valarie Wong completed January 2014 – External examiner for PhD candidate: Valarie Wong, Department of Pathobiology, Ontario Veterinary College, University of Guelph, Canada, Dissertation: *Structure and functions of canine Protein C.*

Priscyla dos Santos Ribeiro completed PhD fall 2023. Dec 2022 – June 2023 visiting PhD Graduate Student. External reviewer and dissertation chapter co-author, Bióloga - Universidade Federal da Bahia (UFBA) Mestra em Microbiologia - PGMICRO UFBA. CAPES Sandwich Doctorate Fellowship, Iowa State University. ISU Sponsor- Dr. Claire Andreasen, co-sponsor USDA ARS Dr. Jarlath Nally. PhD defense September 2023. Dissertation: *Leptospirosis model in hamsters and deer mice; investigative study of biofilm.*

Naima Waheed pending PhD 2024

External Examiner for PhD candidate: Veterinary Science, University of Agriculture, Faisalabad, Pakistan. Dissertation: Epidemiological and pathomorphological investigations of avian salmonellosis in commercial and backyard poultry along with its zoonotic implications.

PROFESSIONAL PRACTICE/SERVICE DUTY

1996-present Cytology and clinical pathology laboratory review
1994-1996 Cytology, clinical pathology laboratory review, and surgical biopsy.
1990-1994 Cytology, clinical pathology laboratory review, surgical biopsy, and necropsy duty.
1986-1990 Surgical biopsy, clinical cytology, and clinical pathology laboratory duty.

REFEREED JOURNAL PUBLICATIONS [*graduate student; +veterinary student mentor]

Andreasen CB. Dietary and medical management of chronic renal failure in dogs and cats. **VM/SAC** 1983;78:62-65.

Andreasen CB, Mahaffey EA. Immunohistochemical demonstration of desmin in canine smooth muscle tumors. **Vet Pathol** 1987;25:211-215.

Andreasen CB, White MR, Swayne DE, Graves GN. Desmin as a marker for canine botryoid rhabdomyosarcomas. **J Comp Pathol** 1988;98:23-29.

Andreasen CB. Neurologic manifestations of ehrlichiosis in a dog. **Mod Vet Pract** 1988;69:96-97.

Andreasen CB, Mahaffey EA, Duncan JR. Intermediate filament staining in the histologic and cytologic diagnosis of canine skin and soft tissue tumors. **Vet Pathol** 1988;25:343-349.

Andreasen CB, Latimer KS, Kirscher IG, Brown J. Protein determination by refractometer and the biuret method in chickens and turkeys. **Avian Dis** 1989;33:93-96.

Andreasen CB, Latimer KS. Separation of avian heterophils from blood using Ficoll-Hypaque discontinuous gradients. **Avian Dis** 1989;33:163-167.

Latimer KS, Kircher IM, Andreasen CB. Separation of turkey heterophils from blood using two-step Ficoll-Hypaque discontinuous gradients. **Avian Dis** 1989;33:571-573.

Andreasen CB, Latimer KS. Cytochemical staining characteristics of chicken heterophils and eosinophils. **Vet Clin Pathol** 1990;19:51-54.

Andreasen CB, Latimer KS. Evaluation of chicken heterophil adherence. **Avian Dis** 1990;34:639-642.

Andreasen CB. What is your diagnosis - idiopathic hemorrhagic effusion. **Vet Clin Pathol** 1990;19:97-98.

Andreasen CB, Mahaffey EA, Latimer KS. What is your diagnosis - thymoma. **Vet Clin Pathol** 1991;20:15-16.

Andreasen CB, Latimer KS, Harmon BG, Glisson JR, Golden JM, Brown J. Heterophil function in healthy chickens and in chickens with experimentally-induced Staphylococcal tenosynovitis. **Vet Pathol** 1991;28:419-427.

Parnell PG, Andreasen CB. What is your diagnosis - branchial cyst. **Vet Clin Pathol** 1992;21:9.

Andreasen CB, Latimer KS. What is your diagnosis - septic meningitis. **Vet Clin Pathol** 1992;21:46.

deHaan JJ, Andreasen CB. Calcium crystal associated arthropathy (pseudogout) in a dog. **J Am Vet Med Assoc** 1992;200:943-946.

Andreasen CB, Carmichael P. What is your diagnosis - *Filaroides hirthi*. **Vet Clin Pathol**, 1992;21:77-78.

Andreasen JR, Andreasen CB. Intestinal adenocarcinoma in a chicken. **Avian Dis**, 1992;36:799-802.

Andreasen JR, Andreasen CB, Anwar M*, Sonn AE. Heterophil chemotaxis in chickens with natural staphylococcal infections. **Avian Dis** 1993;37:284-289.

Andreasen CB, Huber MJ, Mattoon JS. Fibroadenomatous hyperplasia in the mammary gland of a goat. **J Am Vet Med Assoc** 1993;202:1279-1280.

Andreasen CB, Lovelady SD. Evaluation of equine ATIII levels using a chromogenic assay with a fixed band spectrophotometer. **Vet Clin Pathol** 1993;22:42-44.

Andreasen CB. What's your diagnosis - pulmonary carcinoma. **Vet Clin Pathol** 1993;22:67, 87-88.

Andreasen CB, Hedstrom OR, Allison PA. Mediastinal schwannoma in a horse--cytologic, histologic, and immunochemical evaluation. **Vet Clin Pathol** 1993;22:54-57.

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PRESENTATIONS by C. Andreasen:

- May 1986 Andreasen CB: Immunohistochemical staining of canine cytologic and histologic preparations to diagnose soft tissue and subcutaneous tumors. Veterinary Pathology Departmental Research Seminar, Athens GA.
- 1986 Andreasen CB: AFIP Wednesday Slide Seminar Presentations: Lymphocytic cholangiohepatitis and nephritis in a cat.
- December 1986 Andreasen CB: Evaluation of a canine rhabdomyosarcoma with colloid gold immunoelectron microscopy. Course Presentation.
- May 1987 Andreasen CB: Chronic eosinophilic granulomatous pneumonia in a dog. Southeastern Veterinary Pathology Conference, Tifton, GA.
- 1987 Andreasen CB: AFIP Wednesday Slide Seminar Presentations: Pneumonia, eosinophilic granulomatous, chronic.
- November 1987 Andreasen CB: Intermediate filament staining in the cytologic and histologic diagnosis of canine skin and soft tissue tumors. Veterinary Pathology Departmental Research Seminar, Athens, GA.
- November 1987 Andreasen CB, Latimer KS: Slide review - Bacterial meningitis and septicemia. American Society for Veterinary Clinical Pathology. Monterey, CA.
- November 1987 Andreasen CB, Mahaffey EA, Duncan JR: Intermediate filament staining in the cytologic and histologic diagnosis of canine skin and soft tissue tumors. American College of Veterinary Pathologists, Monterey, CA.
- March 1988 Andreasen CB: Immunohistochemical techniques - application to skin and soft tissue tumors of dogs. Resident and intern seminar, Athens, GA.
- May 1988 Andreasen CB, Mahaffey EA: Eosinophilic pustular dermatitis. Sixteenth Annual Southeastern Veterinary Pathology Conference, Tifton, GA.
- July 1988 Andreasen CB, Latimer KS: Separation of avian heterophils from blood using Ficoll-Hypaque discontinuous gradients. Avian Medicine Section/ AAAP American Veterinary Medical Association Meeting, Portland, OR.
- November 1988 Andreasen CB: Separation of chicken heterophils and cell function studies. Veterinary Pathology Research Departmental Seminar, Athens, GA.
- February 1989 Andreasen CB: Neutrophils and cell function studies in man and animals. Resident and Intern Seminar, Athens, GA.
- March 1989 Andreasen CB, Latimer KS: Evaluation of chicken heterophil adherence. Southern Conference on Avian Diseases and Animal Parasites, Athens, GA.
- April 1989 Andreasen CB: Chicken heterophils - isolation and cell function studies. Department of Avian Medicine, Athens, GA.
- May 1989 Andreasen CB, Munsell CL, Prasse KW, Mahaffey EA: Focal cutaneous amyloidosis in a dog. Southeastern Veterinary Pathology Conference, Tifton, GA.
- November 1989 Andreasen CB, Latimer KS, Harmon BG, Glisson JR, Brown J: Evaluation of chicken heterophil adherence in healthy chickens and chickens with staphylococcal tenosynovitis. American Society for Veterinary Clinical Pathology, Baltimore, MD.

- January 1990 Andreasen CB: Evaluation and comparison of neutrophil and heterophil function. Research Seminar, Athens, GA.
- July 1990 Andreasen CB: Heterophil adherence, chemotaxis, phagocytosis and bacterial killing in healthy chickens and chickens with staphylococcal tenosynovitis. Research Seminar, Athens, GA.
- December 1990 Andreasen CB, Latimer KS, Golden JM: Flow cytometric analysis of heterophil phagocytosis in healthy chickens and in chickens with staphylococcal tenosynovitis. American Society for Veterinary Clinical Pathology, Phoenix, AZ.
- July 1991 Andreasen CB, Latimer KS, Golden JM: Flow cytometric analysis of heterophil phagocytosis in healthy chickens and in chickens with staphylococcal tenosynovitis. Avian Medicine Section/AAAP, American Veterinary Medical Association Meeting, Seattle, WA.
- March 1992 Andreasen CB: Avian heterophils and human neutrophils - similarities and differences. Research Seminar, Corvallis, OR.
- May 1992 Andreasen CB: Epizootic bovine abortion. Southeastern Veterinary Pathology Conference, Tifton, GA.
- November 1992 Andreasen CB, Lovelady S: Evaluation of equine ATIII levels using a chromogenic assay with a fixed band spectrophotometer. American College of Veterinary Pathologists, San Diego, CA.
- April 1993 Andreasen CB: Intermediate filaments and tumor diagnosis: An example of test evaluation (specificity, sensitivity, predictive values). Graduate student seminar, College of Veterinary Medicine, Corvallis, OR.
- October 1993 Andreasen CB, Lovelady S. Evaluation of equine AT III levels using a chromogenic assay with a fixed band spectrophotometer. American Association of Veterinary Laboratory Diagnosticians, Las Vegas, NV.
- December 1993 Andreasen CB, Gerros TC, Lassen ED. Evaluation of cellularity, erythroid, and myeloid populations, and stainable iron content in bone marrow from llamas. American Society for Veterinary Clinical Pathology, San Antonio, TX.
- December 1993 Andreasen CB, Rickard L. Cellular and biochemical contents of cisternal and lumbosacral fluid in healthy llamas from Oregon. American Society for Veterinary Clinical Pathology, San Antonio, TX.
- June 1994 Andreasen CB. Clinical chemistry manifestations of toxicologic and nutritional problems in large animals. College of Veterinary Internal Medicine, San Francisco, CA. (Invited)
- June 1994 Andreasen CB. Clinical pathology of llamas-beyond oval red cells. College of Veterinary Internal Medicine, San Francisco, CA. (Invited)
- June 1994 Andreasen CB. Adrenal disease-comparison of horses and companion animals. College of Veterinary Internal Medicine, San Francisco, CA. (Invited)
- June 1994 Andreasen CB. Case studies in large animal renal disease. College of Veterinary Internal Medicine, San Francisco, CA. (Invited)
- August 1994 Andreasen CB. Evaluation of leukocyte function in birds. VIth Congress of the International Society for Animal Clinical Biochemistry; American Society for Veterinary Clinical Pathology Avian Hematology Workshop, Guelph, Canada. (Invited)

- October 1994 Andreasen CB, Snyder SP, Keck TM, Poppenga RH, Braselton WE: Sudden death and hepatopathy associated with elevated hepatic iron levels in Northwestern horses. American Association of Veterinary Laboratory Diagnosticians, Grand Rapids, MI.
- May 1995 Mahaffey, EA, Duncan JR, Andreasen CB (presenter). *Morganella Morganii* enteritis in a Burmese Python. Southeastern Veterinary Pathology Conference, Tifton, GA.
- November 1995 Andreasen CB, Andreasen JR, Thomas JS. Effects of hemolysis on serum chemistry analytes in ratites. American College of Veterinary Pathology, Clinical Pathology Section, Atlanta, GA.
- December 1996 Andreasen C, Pearson E, Smith B, Lassen E, Gerros T. Normal reference ranges and the effects of time and feeding on bile acid concentrations in llamas. American Society of Veterinary Clinical Pathology, Seattle, WA.
- December 1996 Andreasen CB, Newell TK, Gerros TC. Diffuse mesothelioma in a horse. American Society of Veterinary Clinical Pathology, Seattle, WA.
- November 1997 Andreasen C, Jergens A, Hagemoser W. Criteria for gastrointestinal cytology. American College of Veterinary Pathologists (Clinical Pathology Section), Albuquerque, NM.
- November 1997 Andreasen CB, Kramer TT, James MA. Comparison of heterophil phagocytosis for heterophil-adapted *Salmonella enteritidis* (HASE), Wild type *Salmonella enteritidis* (SE), and *Salmonella choleraesuis* (SCS). 78th Conference of Research Workers in Animal Diseases. Chicago, IL.
- March 1998 Andreasen CB, Kramer TT, James MA. Comparison of heterophil phagocytosis for heterophil-adapted *Salmonella enteritidis* (HASE), Wild type *Salmonella enteritidis* (SE), and *Salmonella choleraesuis* (SCS). Western Poultry Disease Conference. Sacramento, CA. Best poster presentation award.
- March 1998 Andreasen CB. Regulation of granulocyte function by ascorbic acid: clinical implications. Microbiology, Immunology, and Preventative Medicine Departmental Seminar. Ames, IA.
- June 1999 Andreasen C, Cowan L, Fenwick B. Erythrocyte morphology in greyhounds with cutaneous and renal vasculopathy. College of Veterinary Internal Medicine, 17th Annual Veterinary Medical Forum, American College of Veterinary Internal Medicine, Chicago, IL.
- June 1999 Andreasen CB. Neutrophil function and clinical application. College of Veterinary Internal Medicine, 17th Annual Veterinary Medical Forum, Chicago, IL. (invited)
- November 1999 Andreasen C, Licklider B, Fulton C. Utilization of a multi-disciplinary resource group for student learning. 50th Meeting, American College of Veterinary Pathologists, Education Section, Chicago, IL.
- June 2001 Andreasen CB. Granulocyte function disorders. Hematopoetic and lymphoid pathology. CL Davis Foundation, University of Illinois, Urbana-Champaign. (Invited)
- July 2001 Andreasen CB, Nelson OL. The use of a plasma D-dimer assay to detect thromboembolic disease in the dog. X International Symposium of Veterinary Laboratory Diagnosticians. Salsomaggiore, Italy.
- November 2001 Andreasen CB, Nelson OL. The use of a plasma D-dimer assay to detect thromboembolic disease in the dog. American Association of Veterinary Laboratory Diagnosticians. Hershey, PA.

- November 2002 Andreasen CB. The cytology of neoplasia. The XVIII PanAmerican Congress of Veterinary Sciences, Havana, Cuba. (invited)
- June 2003 Andreasen CB, Greenlee J. The Continued Emergence of Leptospirosis- Animal Models and Vaccine Development. Seventh Biennial Conference of the Society for Tropical Veterinary Medicine, Vaccine Session, Iguazu Falls, Brazil, 2003.
- July 2003 Andreasen CB, Frank DE, Kinyon J. The effect of ascorbic acid on egg contamination, fecal shedding, and heterophil bacterial killing during Salmonella enteritidis infection. World Poultry Veterinary Association/AAAP. Denver, CO 2003.
- November 2003 Andreasen C, Fales-Williams A, Reynolds D, Ackermann M, Griffith R. A DVM summer scholars program-training future scientists. American College of Veterinary Pathologists and American Society for Veterinary Clinical Pathologists. Banff, CA.
- July 2004 Andreasen C, Frank D. Avian heterophil function and regulation during inflammatory disease. Proceedings of the AVMA/AAAP. Philadelphia, PA.
- November 2004 Andreasen CB, Myers RK. Global perspectives – the veterinary pathologist’s role. American College of Veterinary Pathologists and the American Society for Veterinary Clinical Pathology annual meeting, Orlando, FL.
- June 2005 Andreasen C. Antemortem diagnosis of thromboembolic disease; Utility of D-dimers and FDPs. College of Veterinary Internal Medicine, Annual Veterinary Medical Forum. Baltimore, MD. (invited)
- June 2005 Andreasen C. The platelet and its other life; the role in carcinogenesis and infectious diseases. College of Veterinary Internal Medicine, Annual Veterinary Medical Forum, Baltimore, MD. (invited)
- November 2005 Andreasen C, Sorden S, Roth J, Blanchard T, Metwally S, McKenna T. A digital image base to enhance foreign animal disease education. Proceedings of the American Association of Veterinary Laboratory Diagnosticians 48th Annual Conference. Hershey, PA.
- December 2006 Andreasen C. Thromboembolic disease – The utility of D-dimers and FDPs. Proceedings of the American College of Veterinary Pathologists and the American Society for Veterinary Clinical Pathology. 57th annual meeting, Tucson, AZ. (invited)
- December 2006 Andreasen C, Sorden S, Roth J, Blanchard T, Metwally S, McKenna T. A pathology image data base for foreign, emerging and zoonotic animal diseases. The American College of Veterinary Pathologists and the American Society for Veterinary Clinical Pathology. 57th annual meeting, Tucson, AZ.
- March 2007 Andreasen CB, Roth JA, Suarez DL, Senne DA. Training resources for prevention and intervention strategies during Avian Influenza Outbreaks. 56th Annual Western Poultry Disease Conference, Las Vegas.
- April 2007 Andreasen C, Sorden S, Roth J, Blanchard T, Metwally S, McKenna T. A Pathology Image Data Base for Zoonotic and Emerging Animal Diseases. AAVMC/ASPH Joint Symposium - Partnerships for Preparedness: Future Directions for Schools of Public Health and Colleges of Veterinary Medicine. Atlanta, GA.
- June 2007 Andreasen CB. Using clinical pathology cases to teach clinical pathology and pathology. First Regional Conference on Veterinary Education; 9th Regional Conference in Animal Clinical Pathology and Therapy. Palic, Serbia.
- December 2007 Andreasen CB. A Pathology Image Database for Zoonotic and Emerging Diseases. Iowa Emerging Infectious Diseases Conference, University of Iowa, College of Public Health.

November 2008 Andreasen CB. Granulocyte function and bacterial disease. ISU/UI Meeting for the Vaccine Institute. Newton, IA.

June 2010 Andreasen CB, Roth JA. CEEZAD Education and outreach projects. Education 1st Annual Meeting for Center of Excellence for Emerging and Zoonotic Animal Diseases (CEEZAD). Kansas State University, Manhattan, KS. (invited)

November 2010 Roth JA, Andreasen CB (presenter). Education Consortium, APHIS Veterinary Services Leadership- Panel Discussion. USAHA/AAVLD. Minneapolis, MN. (invited)

February 2011 Roth JA, Andreasen CB (presenter). Overview of education and outreach overlay projects. (invited) Center of Excellence for Emerging and Zoonotic Animal Diseases (CEEZAD) 2011 annual meeting. Florida.

March 2011 Andreasen CB, Wall J. Partnerships to ensure transport of eggs and egg products during a catastrophic disease outbreak. 5th Annual DHS University Network Summit 2011, Washington DC. <http://www.homelandsecurity.org/ReportList.htm>

October 2011 Andreasen CB. Overview of the business continuity plan for the poultry, pork and dairy industries. (invited) FAZD Center Agricultural Screening Tools III, Des Moines, IA.

August 2012 Andreasen CB, Roth JA. Iowa animal agriculture emergency response system, AVMA Meeting, Section: Public and Corporate Practice, Subsection: Current Issues/Advances. San Diego, CA.

April 2013 Andreasen CB, Roth JA. CEEZAD/USDA Higher Education grant: web-based continuing Education courses for veterinarians, graduate students, medical and public health professionals. Education and outreach presentations. CEEZAD Annual Meeting. Nebraska City, NE

March 2014 Andreasen CB. Educational Resources at Iowa State College of Veterinary Medicine; One Health Initiative at Iowa State University, Connecting Education, Research and Outreach. Co-presenters: LK Nolan, Q Zhang. American Association of Veterinary Medical Colleges, Washington DC.

September 2014 Andreasen CB. Developing training programs to support new technology- Panelist. Center of Excellence for Zoonotic Animal Disease Defense (CEEZAD/IIAD) Annual Meeting. Nashville, TN. (invited)

May 2015 Andreasen CB. Department of Homeland Security Education Fair. Washington DC.

October 2017 Andreasen CB. One Health - the Time is Now. 8th International Conference on Animal Health and Veterinary Medicine; One Health - One Society; Toronto, Canada. One Health keynote and session co-chair. (invited)

April 2018 Andreasen CB, Viall A. Webinar case presentations for the American Society for Veterinary Clinical Pathology (ASVCP).

June 2018 Andreasen CB, Roth JA. Insights gained for surveillance and biosecurity from the 2015 High Pathogenicity Avian Influenza (HPAI) outbreaks in the Midwestern U.S., 5th International One Health Congress, Saskatoon, Canada.

November 2019 Andreasen CB, Santos AP, Hodapp A. Fecal *Saccharomyces cerevisiae* resembling other pathogenic yeast in two kittens. American College of Veterinary Pathologists and the American Society for Veterinary Clinical Pathology. 70th Annual meeting, San Antonio, Tx C-05.

- November 2019 Andreasen CB, Hines E+. Comparative hematology - Inappropriate metarubricytosis in canine and human diseases. American College of Veterinary Pathologists and the American Society for Veterinary Clinical Pathology, 70th Annual meeting, San Antonio, Tx. C-01.
- May 2021 Andreasen C. One Health – Our Role. BioConnect Iowa; Vaccines and Immunotherapeutics platform. Virtual meeting, Live Q&A. (invited keynote)
- July 2021 Andreasen C., Moderator NIAMRRE One Health Round Table, NIAMRRE Research Symposium –panelists Drs. Laura Kahn, Bernadette Dunham, Lonnie King, virtual.
- August 2021 Andreasen C, Pineyro-Pineiro P. National Veterinary Student Scholars Symposium. Meet the expert panel-Diagnostic Pathology, virtual.
- November 2023 Andreasen CB, Ngonyama DR, Zamzow J. Panel discussion. Research Hot-Topics: Research Integrity in Practice and How to stay out of trouble, ISU for the College of Veterinary Medicine. (invited)
- April 2025 Andreasen CB, Boggiatto P, Palmer M, Fernandes L VG, Tibbs-Cortes B, Stasko JA, Hamond C, Olsen S, Nally JE, Putz EJ. Peromyscus Deer Mice as an Acute Rodent Model of Leptospirosis. **American Society for Investigative Pathologists (ASIP), Pathobiology Meeting**, Portland, OR, Poster #23, April 2025. (poster presented, unable to attend)

PRESENTATIONS AND PROCEEDINGS FOR FACULTY/GRADUATE ADVANCEMENT AND DIVERSITY

- September 2004 Andreasen CB. How I Did It- Senior Women Faculty. Panel presentation and discussion with J. Bystydzienski and C. Cutrona. Iowa State University (invited)
- March 2006 Andreasen C, Carlson S, MacDonald R, Matibag E, Myers E, Zhu D. Recruiting and retaining a diverse and excellent faculty. 7th annual Iowa State University Conference on Race and Ethnicity (ISCORE), Ames, IA.
- November 2006 Andreasen C, Carlson S, Brooke C, Brumm T. Supporting Flexible Faculty Roles: An Integrated Model for Administrative-Faculty Partnership. Association of American Colleges and Universities, Chicago, IL.
- December 2006 Andreasen C. Work-life issues for academic faculty. Proceedings of the American College of Veterinary Pathologists and the American Society for Veterinary Clinical Pathology. 57th annual meeting, Tucson, AZ. (invited)
- March 2007 Carlson S, Andreasen C (presenter). The Collaborative on Academic Careers in Higher Education (COACHE) survey - faculty satisfaction and experiences related to diversity. 8th Annual Iowa State University Conference on Race and Ethnicity (ISCORE), Ames, IA.
- April 2007 Carlson S, Andreasen C (presenter). Making Progress on a Science and Technology Campus--Policies, Practices, and Assessments. Keeping Our Faculties IV: Recruiting, Retaining, and Advancing Faculty of Color, Minneapolis, MN.
- February 2008 Andreasen CB. Career Flexibility for Graduate Students. University of Florida Spring Symposium on Career Flexibility. Gainesville, Fl. (invited)
- October 2009 Andreasen CB, Wendel J. (Invitation from Provost Hoffman and Associate Provost Carlson). Mentoring and supporting junior faculty: a culture of excellence, collegiality, and collaboration. Iowa State University, Deans and Chairs meeting, Ames, IA.

- April 2012 Andreasen CB. Leadership Ames, Iowa State University Day. Panel representing the ISU Veterinary College. Ames, IA.
- March 2013 Andreasen CB. Career choices and mentorship. ISU Agriculture and Natural Resources/Extension – Women and careers luncheon presentation. ISU, Ames IA.
- September 2014 Andreasen CB. The perfectionist complex- panelist. ISU CVM meeting of the Women’s Veterinary Leadership Development Initiative. Ames, IA. (invited)
- March 2015 Andreasen CB, Blodgett S, Ching B, Constant K, Hayes C. The Graduate Leadership Academy Panel, ISU. (invited)
- January 2016 Andreasen CB, Ching B, Constant K, Strathe M, White P. The Graduate Leadership Academy Panel, ISU. (invited)
- November 2017 Andreasen CB, Cutrona C, Lamont S, Oberhauser A. The Graduate Leadership Academy Panel, ISU. (invited)
- March 2025 Andreasen CB, Gansemer-Topf A, Rosentrater K. Emerging Leaders Graduate College Panel – Mentoring. ISU (invited).

SEMINARS IOWA STATE UNIVERSITY

- February 2004 Andreasen CB. “Animal and Environmental Welfare- The Swedish Perspective.” Animal Science Seminar.
- March 2004 Andreasen CB. “Immunomodulatory Stimulation of Host-Response to *Salmonella enteritidis*.” The Osborn Club, Iowa State University.
- January 2015 Andreasen CB. Working Together to Improve Animal and Human Health, Agricultural Think Tank, Ames, IA (invited)
- April 2017 Barkley M*, Andreasen C. The Effect Humans have on the Raptor Population. Honors Student Poster Presentation. Ames, IA. (*student)
- Nov 2021 Andreasen CB. Opportunities in One Health. CVM Student Public Health Club. Invited.

PRESENTATIONS FOR VETERINARIANS/CONTINUING EDUCATION

- July 1990 Andreasen CB: Diagnostic Cytology for the Practitioner, University of Georgia Continuing Education. University of Georgia, Athens, GA.
- November 1993 Andreasen CB, Gerros TC. Obtaining and interpreting large animal respiratory cytology. Continuing Education Weekend for Veterinarians, Corvallis, OR.
- November 1993 Andreasen CB, Hall, J. Obtaining and interpreting small animal respiratory cytology. Continuing Education Weekend for Veterinarians, Corvallis, OR.
- December 1994 Andreasen CB, Lassen ED. Hematology and cytology of food animals. Annual Fall Conference for Veterinarians, Corvallis, OR.
- December 1994 Andreasen CB, Lassen ED. Hematology and cytology of companion and exotic animals. Annual Fall Conference for Veterinarians, Corvallis, OR.
- June 1995 Andreasen CB. Hematology and clinical pathology of llamas. OVMA Conference, Sunriver, OR.

- June 1995 Andreasen CB. Exotic avian and ratite hematology and cytology. OVMA Conference, Sunriver, OR.
- December 1995 Andreasen CB, Andreasen JR. Exotic avian and ratite hematology, cytology, and necropsy. Third Annual Fall Conference for Veterinarians, Corvallis, OR.
- January 1997 Andreasen CB. Hematology and cytology - cell identification as another piece of the diagnostic puzzle. Iowa Veterinary Medical Association-Veterinary Technician Section, Des Moines, IA.
- June 1997 Andreasen CB. Hematology, cytology, instrumentation, and laboratories-putting it all together to solve the diagnostic puzzle. NDSU Extension Service and NDVMA Continuing Education for Veterinarians, Fargo, ND.
- January 1998 Andreasen CB. Clinical Pathology - In-house versus sending out testing. College of Veterinary Medicine ICN Series, Iowa State University.
- February 1998 Andreasen CB. Erythrocytes and anemias. College of Veterinary Medicine ICN Series, Iowa State University.
- February 1998 Andreasen CB. Bovine leukocytes and platelets . College of Veterinary Medicine ICN Series, Iowa State University.
- March 1998 Andreasen CB. Evaluation of respiratory cytology. College of Veterinary Medicine ICN Series, Iowa State University.
- April 1998 Andreasen CB. Cytology of tumors and lymph nodes. College of Veterinary Medicine ICN Series, Iowa State University.
- April 1998 Andreasen CB. Companion animal leukocyte responses. College of Veterinary Medicine ICN Series, Iowa State University.
- September 1998 Goldman E, Andreasen C. Preparation and cytologic evaluation of fine needle aspiration biopsy specimens. Clinical laboratory, Iowa Veterinary Medical Association Annual Meeting. Ames, IA.
- March 1999 Andreasen CB, Howard M. Clinical pathology-case study. College of Veterinary Medicine ICN Series, Iowa State University.
- April 1999 Andreasen CB. Erythrocyte morphology and mechanisms-understanding the diagnostic puzzle. College of Veterinary Medicine, University of Agriculture, Uppsala, Sweden. (Invited)
- July 1999 Andreasen CB. Getting the most out of your hematology report. College of Veterinary Medicine ICN Series, Iowa State University.
- July 1999 Andreasen CB. Solving the anemia diagnostic puzzle- classifying anemias- what we need to know. Audiovisual presentation. Am Vet Med Assoc, New Orleans, LA.
- July 1999 Andreasen CB. Solving the anemia diagnostic puzzle- how erythrocyte morphology can help. Audiovisual presentation. Am Vet Med Assoc, New Orleans, LA.
- September 2000 Andreasen CB. Urinalysis and cytology. Wet lab. Iowa Veterinary Medical Association-Veterinary Technician Section, Ames, IA.
- September 2001 Andreasen CB. Cytology: Sample collections-staining-slide evaluation. Iowa Veterinary Medical Association 119th Annual Meeting, Ames, IA.

- September 2001 Andreasen CB. Clinical pathology practice tips. Iowa Veterinary Medical Association 119th Annual Meeting, Ames, IA.
- July 2002 Andreasen CB, Troxel M. Cytology of neoplasia in the dog and cat. Audiovisual presentation. Am Vet Med Assoc, Nashville, TN.
- October 2003 Andreasen CB. Case Studies: Solving Diagnostic Dilemmas. Eastern Iowa Veterinary Medical Association, Iowa City, IA.
- September 2004 Andreasen CB. The top ten list- Questions you ask a clinical pathologist. Iowa Veterinary Medical Association 122nd Annual Meeting, Ames, IA.
- June 2007 Andreasen CB. Clinical Pathology Teaching. In association with the 9th Regional Conference in Animal Clinical Pathology and Therapy. Palic, Serbia.
- February 2011 Andreasen CB. Clinical pathology of llamas- beyond oval red cells. Iowa State Camelid Symposium, Ames, IA.

PRESENTATIONS FOR PRODUCER/COMMODITY GROUPS

- March 1995 Andreasen CB. Iron deficiency anemia and iron metabolism in llamas. 8th Annual Llama Herd Health Seminar, Corvallis, OR.
- November 1995 Andreasen JR, Andreasen CB(presenter). What a diagnostic laboratory and clinical pathology laboratory can do for you. Region 4 of the Oregon Emu Association, Keizer, OR.
- March 1996 Andreasen CB. Case studies in clinical pathology. 9th Annual Llama Herd Health Seminar, Corvallis, OR.
- December 1997 Andreasen CB, Roth J, Frank D. Mechanisms of vitamin C in preventing avian staphylococcal osteomyelitis and arthritis. Healthy Livestock Initiative Committee (ILHAC).

FUNDED GRANTS (PI listed first)

University of Georgia

Duncan JR, Mahaffey EA, Andreasen CB (Andreasen wrote grant proposal): Immunohistochemical staining in the cytologic diagnosis of canine skin tumors. Morris Animal Foundation, Englewood, CO. 1986-1987. \$6,535.

Duncan JR, Mahaffey EA, Andreasen CB: Immunohistochemical staining in the cytologic diagnosis of canine skin tumors. Veterinary Medical Experiment Station (VMES), University of Georgia. 1986-1987. \$5,500.

Latimer KS, Andreasen CB, Glisson JR: Clinical Pathology and cell function studies in healthy chickens and chickens with *Staphylococcal tenosynovitis*. VMES Poultry Disease and Research Center, University of Georgia. 1987-1988. \$18,985.

Latimer KS, Andreasen CB, Glisson JR: Clinical pathology and cell function studies in healthy chickens and chickens with *Staphylococcal tenosynovitis*. VMES Poultry Disease and Research Center, University of Georgia. 1988-1989. \$19,858.

Latimer KS, Andreasen CB, Glisson JR: Clinical pathology and cell function studies in healthy chickens and chickens with *Staphylococcal tenosynovitis*. VMES Poultry Disease and Research Center, University of Georgia. 1989-1990. \$22,490.

Oregon State University

Andreasen JR, Andreasen CB: A Comparison of pathogenic and nonpathogenic staphylococcal chemotactic abilities for human neutrophils and avian heterophils. National Institute of Health-Public Health Services, Oregon Research Council, Oregon State University. 1991-1992. \$8,000.

Andreasen CB: A Comparison of avian heterophil and human neutrophil phagocytosis of *Staphylococcus aureus* (capsular types 5 and 8) by flow cytometric analysis. Medical Research Foundation of Oregon. 1992-1993. \$14,772.

Andreasen CB: Flow cytometric analysis of the phagocytosis of *Staphylococcus aureus* (capsular types 5 and 8) by avian heterophils and human neutrophils. National Institute of Health - Public Health Services, Oregon. Research Council, Oregon State University. 1992-1993. \$8,000.

Andreasen CB, Andreasen JR, Sonn AE: Evaluation of avian heterophil function: phagocytosis, killing, and immunomodulating drugs. Oregon State College of Veterinary Medicine 1993-1994. \$6600.

Pearson EG, Craig AM, Andreasen CB, Blythe LL: Perennial ryegrass staggers and atypical pneumonia in Oregon cattle. Oregon State College of Veterinary Medicine 1994. \$6120.

Hall JA, Gradin JL, Andreasen CB: Hepatic secretion of triglyceride: Use of Triton WR 1339 in normal cats and cats with idiopathic hepatic lipidosis. IAMS Company 1994-1995. \$29,142.

Andreasen JR, Andreasen CB: Evaluation of avian heterophil function: bactericidal killing. Oregon State College of Veterinary Medicine. 1994-1995. \$6000.

Andreasen CB, Andreasen JR: The effects of hemolysis on serum chemistry values in broiler & layer chickens and in turkeys. Oregon State College of Veterinary Medicine. 1994-1995. \$6600.

Andreasen CB, Gerros TC: Normal reference ranges of bile acids in llamas and evaluation of the effects of time and feeding. Willamette Valley Llama Association. 1994-1995. \$3600.

Pearson EG, Andreasen CB: Chronic iron toxicity in equidae. Oregon State College of Veterinary Medicine. 1995-1996. \$7350.00.

Andreasen CB, Andreasen JR, Sonn AE: Interaction of heterophils and bacterial killing with immunomodulating drugs. Oregon State College of Veterinary Medicine. 1995-1996. \$6900.00.

Andreasen CB, Andreasen JR, Savage TF: Effects of ascorbic acid on disease manifestations of staphylococcal arthritis and osteomyelitis. Agricultural Research Foundation. 1996-1997. \$6800.00. (Money returned since left institution).

Iowa State University

Roth JA, Andreasen CB, Sorden S: Research position for a newly formed granulocyte research group. HLI Research Support Position Grant. \$27,709 per year. 1996-1999.

Andreasen CB, Roth JA, Frank DE. Mechanisms of antioxidant interactions with avian polymorphonuclear cells and implications for disease prevention. USDA Formula Funds. 1996-1997. \$10,633.

Andreasen CB, Roth JA, Frank DE. Mechanisms of antioxidant interactions with avian polymorphonuclear cells and implications for disease prevention. USDA Formula Funds. 1997-1998. \$14,000.

Kramer T, Andreasen CB. Selection of Salmonella for avirulence and immunogenicity. Healthy Livestock Initiative (HLI) Renewal. \$20,000. 1997-1998. Year 2.

Andreasen CB, Noxon JO, Frank DE. Studies of canine immunocellular function in staphylococcal pyoderma and treatment with immunomodulating drugs. Small Animal Grants Program, ISU. 1997-1998. \$11,000. Year 1.

Andreasen CB, Noxon JO, Frank DE. Studies of canine immunocellular function in staphylococcal pyoderma and treatment with immunomodulating drugs. Small Animal Grants Program, ISU. 1998-1999. \$12,000. Year 2.

Andreasen CB, Ackermann MR, Moon HW. Correlation of Erythrocyte Morphology, Coagulation and Fibrinolysis, and Shiga Toxin Levels in a Bovine Model Utilizing *Escherichia coli* O157:H7 and Implications for Hemolytic Uremic Syndrome. Special Research Initiation Grant Proposal, ISU, 1998-1999. \$10,000.

Andreasen CB, Kramer TT. The use of flow cytometry to determine the pathogenicity of *Salmonella enteritidis*. University Research Grant, ISU, 1998. \$14,992.

Kramer TT, Andreasen CB. Live vaccine against egg-transmission of *Salmonella enteritidis*. Iowa Livestock Health Advisory Council (ILHAC) 2nd year. \$19,000. 1998-1999.

Andreasen CB. Multimedia clinical pathology program: erythrocytes in health and disease. Instructional Developmental Grant, ISU, 1998-1999. \$2000.

Rasmussen L, Andreasen CB (mentor). Role of ascorbic acid in regulating *in vitro* granulocyte (neutrophil and heterophil) bacterial killing. Merck/Merial Summer Veterinary Research Scholars Program. 1998. \$5000.

Rosenbusch RF, Hagemoser WA, Andreasen CB. Characterization of systemic effects produced by *Mycoplasma bovis*. Healthy Livestock Initiative, ISU, 1998-1999. \$20,000. Year 1.

Kline K, Andreasen CB. Differentiation of canine central nervous system diseases producing pleocytosis. Small Animal Grants Program, ISU. 1998-1999. \$10,600. Year 1.

Kline K, Andreasen CB. Differentiation of canine central nervous system diseases producing pleocytosis. Small Animal Grants, Iowa State, 1998-2000. \$11,917, year 2.

Wunn D, Andreasen CB(mentor). Endothelial cell-RBC interactions during hemolytic-uremic syndrome. Graduate/resident research incentive grants. 1999. \$5000.

Bolin CA, Cheville NF, Andreasen CB, Zuerner RL. Development and evaluation of a new trivalent leptospiral vaccine for dogs. Biotechnology Research and Development Corporation. \$72,000. 1999-2000.

Licklider B, Andreasen C, Findlay R, Fowles D, Fulton C, Hendrich S, Huba M, Huiatt T, Reitmeier C, Schimelfenig C, Standly B, Vrchota D, Ware W. Promoting the scholarship of teaching through classroom-based research. Miller Faculty Fellowship. \$25,000. 1999-2000.

Lauer S, Morrison JA, Baldwin CJ, Andreasen CB, Kinyon JM. Subcutaneous totally implantable vascular access systems for feline blood donor populations. Small Animal Research Grant, \$6000, 2000-2001.

Andreasen CB, Bolin CA, Cheville NF, Zuerner RL. Development and evaluation of a new leptospiral vaccine for dogs. Biotechnology Research and Development Corporation. \$75,000. 2000-2001.

Andreasen CB, Nelson L, Ware W. The role of plasma D-dimer to detect pathologic thromboembolic disease in the dog. Morris Animal Foundation. 1999-2001. \$8,210.

Greene S, Andreasen CB (mentor), Baldwin CJ. Comparison of greyhound and non-greyhound platelet counts and platelet function using platelet agonists and shiga toxin. Merck/Merial Summer Veterinary Research Scholars Program. 2000. \$3000.

Andreasen CB, Frank D, Kinyon J. The effect of stress (corticosterone) and ascorbic acid on egg contamination, fecal shedding, and heterophil bacterial killing during *Salmonella enteritidis* infection. ILHAC. \$20,000. 2001.

Greenlee J, Andreasen CB (mentors for Tyson Marshall). Decreased platelet numbers accompanied by gross lesions and other clinicopathologic abnormalities in a guinea pig model of leptospirosis. Merck/Merial Summer Veterinary Research Scholars Program. 2002. \$3000.

Reynolds D, Andreasen CB, Uhlenhopp E. Summer research training of veterinary students. Short-term training for medical students (T35) National Institutes of Health (NIDDK). Continuation 2002-2003 (total \$218,538. 1999-2004.)

Bender H, Danielson J, Mills E, Vermeer P, Andreasen C (collaborator). Creating and distributing a comprehensive on-line learning system for diagnostic reasoning. Department of Education, Learning Anywhere, Anytime Partnership (LAAP)- Fund for the Improvement of Postsecondary Education. 2002-2005, \$959,989.

Mills EM, Bender HS, Danielson JA, Vermeer PJ, Andreasen CB. Teaching Food Animal Diagnostic Skills: A Multi-Institutional Web-Based Approach. USDA-CRESS Higher Education Challenge Grant final year 2004, \$61,208.

Danielson JA, Fales-Williams A, Myers R, Mills EM, Sorden S, Bender H, Andreasen C. Problem solving, communication, critical thinking, and peer assessment: integrating continuous improvement planning into the veterinary pathology curriculum. Miller Fellowship, Iowa State University, 2003-2005, \$25,000.

Andreasen CB, Frank D, Kinyon J. The effect of ascorbic acid on egg contamination, fecal shedding, and heterophil bacterial killing during *Salmonella enteritidis* infection. Midwest Poultry Consortium, 2003-2005, \$25,000.

Cornick N, Andreasen C, Doyle R, Lipert R. Binding and detection of Shiga toxin in blood. ISU University Research Grant. 2003-2004, \$17,343.

Andreasen C, Sorden S, Roth J. A digital image database to enhance foreign animal disease education. USDA Higher Education Challenge Grant. 2005-2007, \$141,262.

Andreasen C, Roth J. Immunomodulatory screening of yeast compounds. Diamond V Mills. 2005-2007. \$85,792.

Palić, D, Andreasen C, Roth J. Enhancing Collaboration and Transfer of Research Methodology with Norwegian National Veterinary Institute and Norwegian School of Veterinary Science, Council on International Programs, Iowa State University, 2006, \$4,708.

Carlson S, Allen B, Andreasen C (co-PI) (Office of the Provost). Flexible faculty careers in the public university-Iowa State University, a change agent for public universities. Alfred P. Sloan Foundation Accelerator Award. 2006-2007, \$25,000.

Ackermann M, Andreasen C. ACVP/STP coalition for PhD dissertation training in anatomic pathology. ACVP/STP Coalition for Veterinary Pathology Fellows. 2006-2009, \$204,905.

Roth J, Andreasen C. Tulathromycin loading and localization in bovine neutrophils and influence on neutrophil function. Pfizer Animal Health. 2006-2008, \$239,825.

Lamont S, Palić,D, Andreasen C. Functional genomics and cellular immunity to Salmonella. NRICGP CSREES-USDA, Animal Genome 43.0, 2007-2009, \$472,346.

Andreasen C, RK Myers. ACVP/STP coalition for residency training in anatomic pathology. ACVP/STP Coalition for Veterinary Pathology Fellows. 2007-2010. \$192,419.

Andreasen C (PI), Collaborators: Millman S, Plummer C, Plummer P, West J. Increasing women's awareness and leadership in global food production. ISU Women's Enrichment Fund Mini-Grants. 2010. \$3120.

Roth JA, Andreasen CB. Subcontract for Education in the Center of Excellence for Emerging and Zoonotic Animal Diseases (CEEZAD); Foreign Animal Disease Education. DHS. \$110,000; 2010.

Roth JA, Andreasen CB, Wall J. Education and Outreach Section, National Center for Foreign Animal and Disease Defense (FAZD), Texas A&M, A decision support system (dashboard) for egg movement. \$25,000; 2010.

Roth JA, Andreasen C, Flaming K. National Center for Foreign Animal and Zoonotic Disease Defense: Enhance and Demonstrate Critical Information for Iowa Animal Agriculture Emergency Response on the FAZD Dashboard TX A&M. \$74,000. 9-1-10 to 6-30-12 Proj # TxAgLife 570696

Simmons H (PI), Roth JA, Andreasen C, Pina M, Hullinger P, Mantelone B. Renewal of the HS- STEM Career Development Program for Foreign Animal and Zoonotic Disease Defense Department of Homeland Security HS-STEM Career Development Grants Program. GRANT10804858 11-60502 Opportunity Number: DHS-11-ST-104-001. 2011 – 2013. \$491,688.00

Roth JA, Andreasen C. Vaccines and Diagnostics for Transboundary Animal Diseases Meeting. DHS 2011-2012 \$182,660 (Direct; \$87,676 IDC; \$270,336)

Roth, J, Mogan J, Flaming K, Andreasen C, Engage to Excel (E2E)—Maintaining Business Continuity and Performing Risk Management during an Animal Disease Outbreak. National Center for Foreign Animal and Zoonotic Disease Defense (FAZD), Texas A&M University Subcontract ISU. \$169,001; total budget \$447,768 (includes IDCs) 7-1-12 to 6-30-14

Roth J PI; Co-PI Andreasen C. DHS Center of Excellence for Zoonotic and Animal Disease Defense - Education and Outreach Component. DHS, Kansas Subcontract. \$1,684,253 (6 yr), 7-1-12 to 6-30-15

Roth J PI, Co-PIs Andreasen C, Hostetter J, Flaming K, Dvorak G. DHS Center of Excellence for Emerging and Zoonotic and Animal Diseases - Education and Outreach Component \$80,000, Kansas State/DHS, 7-1-14 to 6-30-15 (ex 6-30-16) 404-2316.

Mosier D (KSU), Co-PIs: Roth J (ISU), Williams B (AFIP-Path Center), Simmons H (Texas A&M), Richt J (KSU). Collaborators (ISU): Andreasen C, Hostetter J. Curriculum and teaching enhancement for diseases that threaten food security. USDA Challenge Grant. \$198,082; total budget \$713,517(includes IDCs). 9-1-2012 to 9-1-2015

Logue C, Andreasen C. 12 books for 12 women leaders. Women's and Diversity Grant, ISU Office of the Provost. \$1896.28;2014-2015.

Rudolph A (PI), Nusser S, Mogford J, Boehm M (Co-PIs). Building a New Land Grant Ag Biosecurity Consortia to Enhance Pathogen Defense Resiliency. Reecy J (ISU Coordinator), Andreasen C, Roth J (Collaborators/leads animal agro/biodefense), Robertson A (Collaborator/lead plant agrobiodefense), Multi-institutional (CSU, ISU, Texas A&M, UNL), USDA NIFA (Agricultural Biosecurity Coordination Network; Area Code A1181), \$1,000,000 2019-2021. (Not funded)

Nally J (PI), Andreasen CB (collaborator). Palmer M. Foamy macrophages circulating in blood of the hamster model of leptospirosis. KO3 NIH NIAID. Direct costs \$100,000 (NIH is exempt from overhead from ARS). Submitted 2021-2022; not funded.

OTHER PROFESSIONAL ACTIVITIES:

Committees:

University of Georgia

Graduate student application review 1990, Georgia

Oregon State University

Student Computer Laboratory and Usage Committee, 1991-1995

Curriculum Committee, 1993

Computer Task Force 1993

Selection Committee for Director of Instruction and Director of Public Relations 1994

Selection Committee for Pathology Instructor 1994

Biocommunications Selection Committee 1994-1995
Graduate Studies Committee 1994-1996
Legislative Task Force 1995
Selection Committee for Faculty Pathologist 1995
Ad Hoc Scientific Instrumentation Advisory Committee 1995
Ad Hoc Committee Practicing in the State of Oregon 1995
Dean Search Committee, College of Veterinary Medicine 1995-1996
DVM Student Admissions Committee 1996
Selection Committee for Faculty Surgeon 1996

Iowa State University College of Veterinary Medicine

CVM Curriculum Committee 1998-2000
Center for Food Security and Public Health Advisory Board, affiliate member 2007-present
Chair, Search Committee Parasitologist, Veterinary Pathology Department (VPTH) 1999
Chair, Search Committee Clinical Pathologist, Veterinary Pathology Department 2000
CVM Strategic Planning Committee for 2000-2005, 1999-2000
Retreat subcommittee member
CVM Dean Search Committee, member 2000-2001
Biosafety/VDL for necropsy facility 2001-2002
Chair, Search Committee for Chair of Department of Veterinary Microbiology and Preventive Medicine
2002 - 2003
CVM Dean's Search Committee, member 2003 - 2004
CVM Diversity Committee 2005-2009, interim chair.
Panels to interview prospective veterinary students 2007, 2009, 2011-2017- backup
Chair, Search Committee for public relations/communications 2013
Lead, Develop AVMA COE Mock site visit document, 2016
Ex-officio member as per Associate Dean 2010-2016
Admissions Committee
Curriculum Committee
Academic Standards Committee
Computer, Library and Information Management
Student- Faculty Relations Committee
AVMA-COE Accreditation Task Force, 4 member lead team, 2016-2017
Safety website development
Ad hoc committee, under-represented student recruitment, member, 2016-2017
Ad hoc committee, reducing cost and value added to DVM, member, 2016-2017
Search committee clinical pathologist, VPTH, member, 2019
Search committee anatomic pathologist, VPTH, member, 2019
CVM Admissions, member 2017- 2020
Subcommittee lead for early admissions information, 2018
Review committee, Talbot Professorship, 2020
CVM Search committee, VCS radiation oncology, member 2021
CVM Search committee, VCS medical oncologist, member 2021
CVM Academic Standards Committee, member 2017- 2022
CVM Faculty Senate Caucus Chair, 2018 – 2021; member 2021-2022
Department of Veterinary Pathology Faculty Senate representative 2016-2022
CVM Diversity and Inclusion Committee, member, 2017-2023
Subcommittee for Office of D&I, 2017-2018.
CVM Frederick Douglass Patterson Committee 2022-2023
Department of Veterinary Pathology Promotion and Tenure Committee 2011-2024
Department of Veterinary Pathology Director of Graduate Education 2023-2024
CVM Veterinary Student Advisor 1996-2024
CVM Art Committee, member 2016-2024
CVM Term Faculty Advancement Committee, member 2019-2024
Interim Chair 2020-2021
Term Advancement Workshop Panel for CVM Dec 2023
CVM Faculty, Staff and Alumni Awards 2023-2024
Department of Veterinary Pathology Graduate Curriculum Committee 1997-present

Iowa State University

Ad Hoc University Committee for Curriculum Development in U.S. Diversity and International Perspectives Position 1997

Interdepartmental Graduate Program, Immunobiology

Admissions Committee, 1999-2000

Supervisory Committee, 2000-2003

University Taskforce on the Recruitment and Retention of Women and Minority Faculty 2002-2003

Strategic Planning Committee for Iowa State University 2004-2005

Provost's Task Force Reviewing the Faculty Mentoring Program, Chair 2006

Committee for ISU Foundation Departmental Leadership Award 2006

Member, Search Committee for Wildlife Biologist, NREM 2006

Iowa State University Women's Leadership Group, 2001-2010

Women's Leadership Consortium, Group leader, 2006

ISU Department Chair Cabinet/Council 2001-2007

Keeping Our Faculty of Color Leadership Team 2005-2006

University Committee for Women, member, 2006- 2011

ADVANCE Symposium Committee 2007-2008

National Campaign Committee/Campaign ISU 2007-2010

Faculty Responsibilities Work Group 2009

ADVANCE Council 2011

CVM Representative Search Committee Member, Director of Reiman Gardens 2014

Safe Zone Training 2014

Women Impacting ISU Calendar Selection Committee for 2016 calendar, member

Research Colloquium Committee 2016, member

ISU Student On-Boarding Committee, Subcommittee chair, 2016-2018

ISU Campus Climate Survey Faculty Workgroup, member 2018

ISU Child Care Task Force, member 2019-2020

ISU Search Committee for National Institute of Antimicrobial Resistance Research and Education

(NIAMRRE), for Associate Director of Operations and Associate Director of External Relations 2020

ISU Review Committee for Cost-Sharing Research instrumentation (COSRIF), member 2016-2021

ISU Vice President for Diversity and Inclusion Council, member 2015- 2021

The Leadership Imperative, Office of Diversity and Inclusion 2017, contributor

ISU Faculty Senate

Council on University Resource Policies and Allocations, member, 2016-2017

Faculty Development and Administrative Relations Council, Chair, 2017-2021, member 2021-2022

Executive Board, member, 2017-2021

Administrative Committee, member, 2017- 2021

Representative Committee, member, 2018-2021

CVM Caucus Chair, 2018-2021; member 2021-2022

VPTH representative – 2016-2022

ISU graduation ceremony, Faculty Marshal, to replace spring 2020 graduation, Oct 2021

ISU Strategic Plan Working Group 2021-2022

ISU Publication Subvention Review Committee, member 2019-2022

ISU Residency Appeals Committee, member 2010-2022

CVM Dean Review Committee 2023

University Museums Advisory Committee, member 2015-2023

ISU Faculty Senate Honorary Degree review committee 2023-2024

ISU Mid-career Research Award Committee 2023- 2024

ISU University Professor Committee 2024

ISU CVM Dean Review Committee 2023

ISU CALS Dean Review Committee 2023-2024

ISU Non-Academic Use of Animals Committee, member 2014-2025

Per past Associate Dean position:

Undergraduate Program Council 2010-2016

Academic Associate Deans 2010-2016

Council for International Programs 2012- 2016

Associate Deans- Credit Distance Education 2010--2016
ISU IT Advisory Committee 2013- 2015; now Information Technology Leadership Committee (ITLC),
2015- 2016

ISU current:

University Museums Advancement Council, member 2018-present
National Institute of Antimicrobial Resistance Research and Education (NIAMRRE), member 2018-
present

Federal Agency

Search Committee National Animal Disease Center Director, Member. 2005-2006.
National - listed under organizations

Journal Reviewer:

Reviewer **Advances in Small Animal Medicine and Surgery** 1991-1992.

Ad Hoc Reviewer **Journal of Zoo and Wildlife Medicine** 1996-2011

Ad Hoc Reviewer **Veterinary Pathology** 1991-present.

Ad Hoc Reviewer **Journal American Veterinary Medical Association** 1993-present.

Ad Hoc Reviewer **Journal of Veterinary Diagnostic Investigation** 1994 - present.

Ad Hoc reviewer **American Journal of Veterinary Research** 1993-present.

Ad Hoc Reviewer **Veterinary Clinical Pathology** 1994-present.

Ad Hoc Reviewer **Avian Pathology** 1997-present.

Ad Hoc Reviewer **Poultry Science** 2019- present

Editorial Board, **Comparative Haematology International** (changed to **Comparative Clinical Pathology**)
1996- 2009, 2016-present.

Frontiers Veterinary Science 2016-2021; Review Editor Veterinary Humanities and Social Sciences 2016-2020;
Veterinary Experimental and Diagnostic Pathology 2020-2021.

Dr. William G. Van Alstine, DVM, PhD, received his DVM from the University of Missouri, College of Veterinary Medicine in 1981. After working in a predominantly large animal practice in Minnesota, he attended Iowa State University and received a PhD in anatomic pathology in 1987. He was board-certified by the American College of Veterinary Pathologists and joined the faculty of Purdue University College of Veterinary Medicine and Indiana Animal Disease Diagnostic Laboratory that same year. Dr. Van Alstine retired as Professor of Veterinary Pathology (Emeritus) from the College of Veterinary Medicine, Purdue University in 2014 after 28 years at Purdue. During his tenure at Purdue University, he served as a diagnostic and research pathologist and helped teach/mentor over 100 pathology graduate students/residents and approximately 2000 veterinary students. His academic interests focused on animal models for large animal diseases, investigating and defining induced and naturally occurring toxic and infectious diseases of animals, and developing animal models for non-clinical human medical device testing. He has over 125 publications (scientific journals and book chapters). At Purdue, he held multiple administrative positions, including Head of Pathology in the ADDL and the Department of Comparative Pathobiology, Associate Director of the ADDL, and Chairman of the Purdue University Animal Care and Use Committee. After retiring from Purdue University, Dr. Van Alstine joined Cook Medical, a global medical device company, and served as Director of Veterinary Pathology until 2022.

Dr. Van Alstine has been a member of AAVLD for 42 years and has served on many/most administrative, program, and scientific committees. In 1997, he had the privilege to serve as President of the AAVLD. Over the past 42 years, he has been continually amazed at the progress of diagnostic medicine in the United States and equally amazed at how veterinary diagnostics worldwide are more advanced and increasingly more available in all parts of the world, including developing countries. That accomplishment is due in a very large part to the efforts of the AAVLD. He considers it a privilege to have been a part of this wonderful scientific group!

Since 2022, he has still occasionally provided pathology and regulatory support to medical device companies and interacts with the FDA and regulatory bodies around the world. More importantly, he enjoys traveling with his wife, Patti, to visit their 4 adult children and (currently) 13 grandchildren. He plans to complete the final leg of a multi-year section hike of the Appalachian Trail in 2026.

Robert H. Poppenga, DVM, PhD, received my DVM and PhD degrees from the University of Illinois in 1978 and 1987, respectively. During my PhD studies, I was also a staff veterinarian for the National Animal Poison Control Center at the College of Veterinary Medicine (the center subsequently became the ASPCA Animal Poison Control Center). Since receiving my PhD and residency training in veterinary toxicology, I have held faculty positions at Michigan State University, University of Pennsylvania, and University of California at Davis (UCD) veterinary schools. I am currently a Full Professor, Emeritus, having been a veterinary toxicologist at the California Animal Health and Food Safety Laboratory System (CAHFS), School of Veterinary Medicine, UCD, from 2004 to 2025. I retired from CAHFS on July 1, 2025.

I have nearly 35 years of experience in veterinary toxicology and food safety as a result of working in several veterinary diagnostic laboratories. I have routinely conducted forensic investigations into a variety of potential accidental and malicious animal poisonings involving numerous species, often involving an assessment of causation. I have supervised and directed state-of-the-art veterinary diagnostic toxicology sections with a broad array of analytical capabilities and diversity of case submissions.

I have taught professional, graduate, and undergraduate students using a variety of teaching formats, including case- and problem-based learning modalities. I have also trained residents in veterinary toxicology as part of my duties with CAHFS. I initiated a bi-weekly collaborative case-based discussion group with veterinary toxicologists and residents at the University of Kentucky, Iowa State University, and Kansas State University. While at the University of Pennsylvania, I led the effort to develop a computer-aided learning module on poisonous plants. As an Emeritus faculty member, I continue to serve on the Admissions and Graduate Student Advisory Committees of the Forensic Science Graduate Group and the Graduate Student Advisory Committee of the Pharmacology and Toxicology Graduate Group at UCD.

In my role as Toxicology Section Head at CAHFS, I supervised 11 FTE analytical chemists and staff with over 100 years of combined experience across all analytical systems. The Toxicology Section has a full array of analytical capabilities, including LC/MS (and HRMS), GC/MS, ICP/MS, LC/ICP/MS, and ELISA analytical platforms. The Section has a unique capability to approach chemical contamination incidents from a variety of clinical and analytical perspectives. My research interests have included diagnostic veterinary toxicology, wildlife toxicology, and the development of biomarkers for chemical exposure. I am an author/co-author on more than 160 peer-reviewed publications and numerous book chapters.

Since 2004, the CAHFS Toxicology Section has received support through the US Food and Drug Administration's Food Emergency Response Network (FERN) and Veterinary Laboratory Investigation and Response Network (VetLIRN) Chemistry programs, and I have been the PI on those cooperative agreements.

I am active in several professional organizations including the American Association of Veterinary Laboratory Diagnosticians (Regional Executive Board Member, Journal of Veterinary Diagnostic Investigation Editorial Board, and member of the Committee on Toxicology and Environmental Issues), American Board of Veterinary Toxicology (Secretary/Treasurer, Vice President, President, Examination Committee Chair, and Education Committee), and the American Veterinary Medical Association (Committee on Environmental Issues).

December 18, 2025

François Elvinger, Dr.med.vet., Ph.D., dipl. ACVPM & ECVPH

fe65@cornell.edu tel: +1 607 882 0057
Animal Health Diagnostic Center
240 Farrier Road
Ithaca, NY 14853, U.S.A

Biographical Sketch

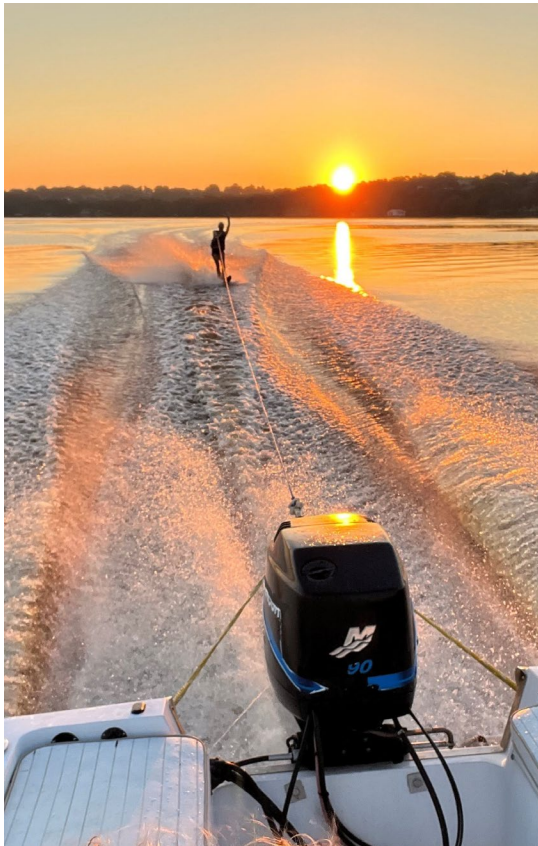
François Elvinger, Professor of Veterinary Epidemiology, Associate Dean for Diagnostic Operations and Government Relations, and Executive Director of the Cornell University College of Veterinary Medicine Animal Health Diagnostic Center and New York State Veterinary Diagnostic Laboratory, obtained his veterinary and Dr.med.vet. degrees at the Hannover Veterinary School in Germany (1981 and 1983), his Ph.D. from the University of Florida (1990), before taking his first faculty position as veterinary epidemiologist at the University of Georgia Veterinary Diagnostic and Investigational Laboratory in Tifton, Georgia (1990-1997), then moving on to the Large Animal Clinical Sciences Department and Production Management Medicine group at the Virginia-Maryland College of Veterinary Medicine at Virginia Tech in Blacksburg, Virginia (1997-2015). He developed the Virginia Tech Public Health Program with the first veterinary college based MPH degree program and the Population Health Sciences Department, of which he was the founding director and founding head, before joining the Cornell Animal Health Diagnostic Center in 2015.

After joining the AAVLD and the USAHA in 1991, he chaired his first AAVLD committee in 1994, the Committee on Animal Disease Reporting, which produced the DxMonitor, an early veterinary diagnostic laboratory-based animal health and disease quarterly report. He has held leadership positions as chair, founding chair, and co-chair of committees of both organizations ever since, with his colleagues, mentors, and close friends Drs. Bruce Akey, Mo Salman, and Mark Thurmond, including co-founder of the AAVLD Epidemiology Committee and of one of the two earliest AAVLD and USAHA Joint Committees, on Animal Disease Surveillance and Animal Health Information, now Animal Health Surveillance and Information Systems, which he both co-chaired for more than 10 years. He has co-chaired the Steering Committee on the National Animal Health Reporting System (NAHRS; 1998–2011) and has chaired the Steering Committee of the National Animal Health Surveillance System (NAHSS; 2004–2012), both joint projects of AAVLD, USAHA, and USDA:APHIS:VS and CEAH. His work on launching NAHRSS and NAHSS earned him the APHIS Administrator's Award in 2007 at the annual meeting in Reno, NV. Dr. Elvinger held the AAVLD Presidency with its many pre- and post-presidency chair commitments in 2015. He served as a member of the Coordinating Council of the National Animal Health



Laboratory Network (NAHLN), and now co-chairs the Joint Committee on the NAHLN, as well as, again, the AAVLD Epidemiology Committee.

He is a diplomate of both the American College of Veterinary Preventive Medicine (1994) and the European College of Veterinary Public Health (2005), with research, teaching and service interests in surveillance methodology for high impact disease and validation of diagnostic tests in particular for infectious, including zoonotic diseases.



Awards

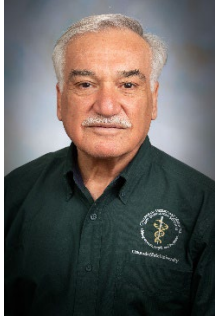
2007 USDA APHIS Administrator's Award for Animal Health
2009 USDA APHIS VS Deputy Administrator CVO Certificate of Appreciation
2022 American Association of Veterinary Laboratory Diagnosticians, **Pope Award**

AAVLD Workshops etc.

Editorial Review Group: Dx Monitor (AAVLD/USAHA/USDA:APHIS:VS) (1992-1997)
"Identification and Consolidation of Existing Data Sources and Standardization of Disease Definitions and Reporting" workshop (1995), coordinator, organizing group
Workshop on Validation of Veterinary Diagnostic Tests. Co-organizer. American Association of Veterinary Laboratory Diagnosticians, Annual Meeting. October 17, 2002. St. Louis, Missouri
Special Session on Validation of Veterinary Diagnostic Tests. Co-organizer (with Dr. Mark Thurmond). American Association of Veterinary Laboratory Diagnosticians. Annual Meetings. 2003, San Diego, California; and 2004, Greensboro, North Carolina
Criteria for Validation of Diagnostic Tests. (Elvinger F and Thurmond MC). American Association of Veterinary Laboratory Diagnosticians, Special Session on Validation of Veterinary Diagnostic Tests for High Impact Foreign Animal Disease (Foot and Mouth Disease, Classical Swine Fever, Exotic Newcastle Disease). San Diego, CA. October 11, 2003
Workshop on Sampling in Veterinary Diagnostic Laboratories. Co-organizer (with Drs. Mark Thurmond and Claudia Munoz-Zanzi). American Association of Veterinary Laboratory Diagnosticians, Annual Meeting. October 12, 2006. Minneapolis, Minnesota
Veterinary Diagnostic Laboratory Epidemiology Toolbox – (with Drs. Suzanne Burgener and Albert Rovira) Course offered at and extended from the American Association of Veterinary Laboratory Diagnosticians. Annual Meeting. October 9, 2009. San Diego, California

AAVLD / USAHA Committee chair / co-chair

Committee on Animal Disease Reporting, American Association of Veterinary Laboratory Diagnosticians (1993-1997), Chair (1994-1997)
Subcommittee on Standard Operating Procedures for Committee Chairs and Committees, Chair, Program Committee, United States Animal Health Association (1999-2004)
Epidemiology Committee, American Association of Veterinary Laboratory Diagnosticians. Founding co-chair (2000-2010, **2023-present**); Epidemiology Scientific Session, Annual Meeting, American Association of Veterinary Laboratory Diagnosticians. Moderator (2000-2006, 2008, 2025)
Joint-Committee on Animal Health Information Systems, United States Animal Health Association and American Association of Veterinary Laboratory Diagnosticians, Co-chair (1998-2009); then Joint-Committee on Animal Health Surveillance and Information Systems (AHSIS), Co-chair (2009-2011)
Steering Committee of the National Animal Health Reporting System (NAHRS), a joint project of the USAHA, AAVLD, and United States Department of Agriculture:Animal Plant Health Inspection Service:Veterinary Services, Co-chair (1998-2011)
National Animal Health Reporting System (NAHRS) Unified Methods and Rules, effective January 1, 2001. Authored by the NAHRS Steering Committee (Akey BL, Elvinger F, co-chairs)
Strategic Plan, National Animal Health Surveillance System. USDA:APHIS:VS:CEAH:National Center for Animal Health Surveillance. December 2004. Authored by the National Animal Health Surveillance Systems Steering Committee (Elvinger F, chair)
Steering Committee of the National Animal Health Surveillance Systems (NAHSS). United States Department of Agriculture:Animal Plant Health Inspection Service:Veterinary Services:National Center of Animal Health Surveillance, Chair (2004-2009); sub-Committee on NAHSS of AHSIS Committee, Co-chair (2009-2012)
American Association of Veterinary Laboratory Diagnosticians, Vice President (2012-13); President Elect (2013-14); President (2014-15); Past President (2015-16)
American Association of Veterinary Laboratory Diagnosticians, Awards Committee, Chair (2015-16), NE-AAVLD, Convener of 1st meeting in Atlantic City, NJ (2017); Chair of Organizing Committee, Newport, RI (2018); co-organizer, Annapolis, MD (2019)
National Animal Health Laboratory Network (NAHLN), Coordinating Council member (2018-2022)
Joint-Committee on the NAHLN, United States Animal Health Association and American Association of Veterinary Laboratory Diagnosticians, Co-chair (2022-present)



Dr. Mo Salman, DVM, MPVM, PhD, DACVPM, FACE, Hon. Dipl. AVOHS, is a distinguished professor in the Department of Clinical Sciences at Colorado State University (CSU) and the founder and director of the Animal Population Health Institute within the College of Veterinary Medicine and Biomedical Sciences. With over 40 years of experience, Dr. Salman has an extensive educational background in veterinary medicine (BVMS), preventive veterinary medicine (MPVM), and comparative pathology (PhD). He is a Diplomate of the American College of Veterinary Preventive Medicine (ACVPM), a Fellow of the American College of Epidemiology (ACE), and an Honorary Diplomate of the American Veterinary One Health Society (AVOHS).

Dr. Salman's research specializes in surveillance and survey methodologies for animal diseases and veterinary public health, with a focus on infectious diseases and zoonoses. He is the editor of the 2003 book *Animal Disease Surveillance and Survey Systems: Methods and Applications*. Over the past 30 years, he has led training sessions and workshops in epidemiology, surveillance, disease management, and risk assessment, sponsored by various local, national, and international organizations.

Dr. Salman has made significant contributions to stabilizing and reconstructing national animal and public health programs in countries such as Bosnia, Afghanistan, Iraq, Mexico, East Africa, Georgia, Turkey, and Armenia. He has worked to enhance technical human infrastructure and border security to mitigate the spread of human and animal diseases. Additionally, he has advised on plans to build national biosecurity for animal health and develop public health strategies in Uruguay, Saudi Arabia, Iraq, Georgia, Armenia, and Kuwait.

Dr. Salman served as chair of the initial WOAHA (formerly OIE) committee that established guidelines for BSE (bovine spongiform encephalopathy) country status and was a member of the first European Union committee assessing BSE geographical risks among EU and non-EU nations.

From 2009 to 2018, Dr. Salman was Editor-in-Chief of Elsevier Preventive Veterinary Medicine. In 2013, he served as a Jefferson Science Fellow with the U.S. Department of State, acting as a scientific advisor to the Africa Bureau. From 2015 to 2016, he coordinated the U.S. Department of State Biosecurity Engagement Program in Afghanistan, focusing on securing points of entry to prevent the introduction of highly pathogenic human and animal diseases. He has also served for the last nine years as a

NATO civil expert on resilience and as a member of the WHO Joint External Evaluation (JEE) team.

Currently, Dr. Salman is dedicated to education, developing case studies for field training of public animal health officers worldwide and contributing to CSU's academic programs for undergraduate, graduate, and professional students. His impactful contributions to animal and public health systems globally have cemented his reputation as a leader in his field.

Dr. William Wilson, Ph.D., obtained his BSc (honors, 1979) and PhD (1985) in Animal Science at the University of Illinois. He was a Cromwell Postdoctoral Fellow before joining the USDA, ARS, Arthropod-Borne Animal Diseases Research Laboratory (ABADRL), located in Laramie, WY, as a Research Microbiologist in 1986. Dr. Wilson was relocated to Manhattan, KS, in 2020 with the Arthropod-Borne Animal Diseases Research Unit (ABADRU). He transferred to the Foreign Arthropod-Borne Animal Diseases Research Unit (FABADRU) at the National Bio and Agro-defense Facility (NBAF) in 2020 before retiring in December 2024. He is a Marty Vanier and Bob Krause Biosecurity Research Institute Fellow and has served as a scientific consultant to the FAO/IAEA Joint Animal Health Division, as well as an Affiliate Faculty at Colorado State University, Texas A&M and Washington State University. He currently serves as Adjunct Professor at Kansas State University and an Extraordinary Lecturer at the University of Pretoria, South Africa. He has been a member of several professional societies including AAVLD since 1999 and continues to serve as the Membership Committee Chair for the World Society of Virology. Dr. Wilson's research focuses on the molecular biology, pathogenesis and diagnostics of arboviruses that cause important transboundary animal diseases (TADs) including bluetongue, Japanese encephalitis, Rift Valley fever and Vesicular Stomatitis. His work involves collaborative projects with national and international partners, including scientists from academia, government, and the bioscience industry in countries such as the US, Australia, Africa, and Japan. Recent collaborative partners include scientists from Kansas State University, the Canadian Food Inspection Agency, the Onderstepoort Veterinary Institute and the University of Pretoria in South Africa, as well as, in Kenya, the International Livestock Research Institute, the Kenya Agricultural Livestock Research Organization, the Kenya Department of Veterinary Services, and the Kenya Wildlife Services. Dr. Wilson's scientific activities have generated diagnostic tools and vaccines for the livestock industries as well as 10 book chapters and more than 150 peer-reviewed manuscripts.

Amy K. Swinford, DVM, MS, Diplomate (Emeritus) ACVM, is director of the Texas A&M Veterinary Medical Diagnostic Laboratory. Dr. Swinford is a four-time graduate of the University of Illinois, earning bachelor's degrees in both microbiology and veterinary science, a DVM, and a master's degree in veterinary pathobiology. She became a diplomate of the American College of Veterinary Microbiologists in 1997. She started her career as a diagnostic virologist at the University of Nebraska Veterinary Diagnostic Center in Lincoln, studying pseudorabies virus and what was then a recently discovered disease, porcine reproductive and respiratory syndrome (PRRS). During this time, Dr. Swinford also served as a public health officer in the United States Air Force Reserve.

In 2005, Dr. Swinford joined the Texas A&M Veterinary Medical Diagnostic Laboratory (TVMDL) as section head of diagnostic bacteriology/mycology. She also enjoyed collaborative research, and serving as an adjunct assistant professor in the College of Veterinary Medicine & Biomedical Sciences at Texas A&M University, where she delivered lectures in veterinary microbiology and infectious disease to first through third-year veterinary students.

Dr. Swinford became TVMDL's microbiology branch chief in 2012, overseeing the bacteriology, virology, serology, and molecular diagnostics sections. As branch chief, she provided leadership, oversaw stringent quality assurance and safety programs, and implemented new testing protocols. She became particularly interested in the quality assurance standards that applied to veterinary diagnostic laboratories, which led her to join the AAVLD Accreditation Committee's Auditor Pool, and ultimately the AAVLD Accreditation Committee. She enjoyed many years serving on the Committee, auditing AAVLD laboratories, and seeing the AAVLD Accreditation Program Standard in action as it enhanced the quality of veterinary diagnostics across the United States.

Dr. Swinford became associate director of TVMDL in 2015 and was appointed TVMDL's sixth agency director in 2021. During her tenure, Dr. Swinford coordinated TVMDL's responses to numerous disease outbreaks such as vesicular stomatitis, contagious equine metritis, anthrax, chronic wasting disease, highly pathogenic avian influenza, and SARS-CoV-2. She helped implement TVMDL's testing of human patients for SARS-CoV-2 during the COVID-19 pandemic and guided her laboratory's response to the outbreak of highly pathogenic avian influenza in dairy cattle, first discovered in the Texas Panhandle in 2024.

Dr. Swinford has been recognized with awards from the Texas A&M University System, including the TVMDL Director's Excellence Award, three Vice Chancellor's Awards in

Excellence, and the Texas A&M System Regents Fellow Service Award. On November 13, 2025, the Texas A&M System Board of Regents bestowed the title of director emeritus in recognition of Dr. Swinford's 20-plus years of dedicated service to TVMDL and its clients.

Dr. Swinford has been an active member of AAVLD since 1992, attending her first AAVLD/USAHA Annual Meeting in 1993 in Las Vegas, Nevada. She's had the privilege of serving on numerous committees, including Accreditation, Subcommittee on Bacteriology and Mycology, Subcommittee on Antimicrobial Susceptibility Testing, Membership, Government Relations, Laboratory Directors, Strategic Planning, AAVLD/USAHA Joint Committee on the NAHLN, AAVLD Committee Chairs and Program Committee, AAVLD House of Delegates, and the AAVLD Executive Board.

Alexis Thompson, DVM, PhD, DACVPM (Epidemiology), is the Resident Director of the Texas A&M Veterinary Medical Diagnostic Laboratory (TVMDL) in Canyon, TX. She first joined TVMDL in 2023 as a bovine veterinary diagnostician and was promoted to resident director in 2024. In 2015, Thompson earned a bachelor's degree in animal science from the University of Kentucky. She earned a Doctor of Veterinary Medicine degree in 2019, completed a residency in Population and Preventive Medicine in 2022, and earned her doctorate in 2023, all from Mississippi State University. Thompson is boarded in veterinary preventive medicine (2023) and epidemiology (2024).

Mike Filigenzi, BA, served as the manager of the Toxicology Laboratory at the California Animal Health and Food Safety Laboratory when he retired in 2023. Mike graduated with a Bachelor's Degree in Psychobiology from Pitzer College and shortly thereafter began working in the then-nascent contract environmental laboratory industry. Mike gained extensive experience in various mass spectrometry techniques over 20 years as a chemist, supervisor, and manager in environmental laboratories. In 2000, Mike joined the Equine Analytical Chemistry Laboratory at CAHFS as a staff chemist. In 2002, he moved to the Toxicology section as a supervisor and has been there since, moving into the lab manager position in 2021. Over the years, Mike has specialized in the development of analytical methods using various types of chromatographs and mass spectrometers. He is a tremendous nerd and believes that mass spectrometers are pretty much the coolest things ever.

Dr. Charnas, DVM, MPH, holds a Doctor of Veterinary Medicine (DVM) degree from the University of Florida College of Veterinary Medicine and a Master of Public Health (MPH) degree from the University of Florida School of Public Health & Health Professions. She is currently serving as a third-year veterinary toxicology resident at the Kansas Veterinary Diagnostic Laboratory, where her responsibilities include interpreting diagnostic cases and educating third and fourth-year veterinary students.

My name is Brooklyn Cauwels and I am a Diagnostic Associate II in the Serology section of the Iowa State University Veterinary Diagnostic Laboratory. I received a BS in Biology, minor in Chemistry from ISU and have completed my second year of graduate school at ISU, pursuing a MS in Immunobiology. Since starting in my position, I have learned many serological assays across multiple testing platforms such as ELISAs, Luminex FMIA, IFAs and agglutination assays. I was a major technician in the Covid-19 Public Health Testing Services lab during the peak of the pandemic, and collaborated with fellow technicians to develop and implement a sample processing method to mitigate the risk of contamination of samples for Health Assurance Testing Service clients. Additionally, I am the primary technician on the blocking NP ELISA in response to the emergence of HPAI in dairy cattle, and have worked on multiple projects related to the validation of assays and specimens for use in the Serology section at the ISU VDL. For my research, I have been trained in PBMC isolation, cell isolation, and ELISpot assays, cell proliferation assays, flow cytometry as well as extraction and PCR. This has enabled me to expand my knowledge and capabilities, aligning with my goals to work in research related to diagnostic techniques, infectious diseases, and host-pathogen interaction at a more advanced level.

Chloe Stenkamp-Strahm has a DVM and PhD from Colorado State University, and a research background studying ruminant herd health and infectious diseases. Her post-doctoral work at Colorado State University focused on measuring impacts of the highly pathogenic avian influenza H5N1 virus in U.S. dairy cows, including studying patterns of herd and individual animal level detection of virus over time. Chloe has recently become an Assistant Professor and Extension Veterinarian at Utah State University, where she teaches veterinary public health and continues research on H5N1 and other zoonotic diseases within the state of Utah.

Bala Mounika Reddi is currently a graduate student in Dr. Rahul K. Nelli's laboratory at Iowa State University. Her research centers on advancing molecular diagnostic workflows in veterinary diagnostic laboratories, with an emphasis on high-throughput RT-qPCR platforms and automation for large-scale detection of animal pathogens. She is particularly focused on developing rapid, precise, and cost-effective diagnostic solutions to strengthen disease surveillance and support modern veterinary diagnostic needs.

Sree Varsha Kuduva Sankar Babu (Varsha Sankar) is a graduate student pursuing a Master's degree in Comparative Biomedical Sciences at the College of Veterinary Medicine, Oklahoma State University. She holds a degree in Veterinary Medicine and a Postgraduate Diploma in Veterinary Laboratory Diagnosis, with professional experience in veterinary clinical pathology. Her research focuses on the isolation and characterization of bacteriophages targeting multidrug-resistant bacterial pathogens, including *Pseudomonas.aeruginosa* and *Salmonella.enterica*, conducted under the guidance of Dr. Akhilesh Ramachandran, ACVM diplomate.

Dr. William Wilson, Ph.D., obtained his BSc (honors, 1979) and PhD (1985) in Animal Science at the University of Illinois. He was a Cromwell Postdoctoral Fellow before joining the USDA, ARS, Arthropod-Borne Animal Diseases Research Laboratory (ABADRL), located in Laramie, WY, as a Research Microbiologist in 1986. Dr. Wilson was relocated to Manhattan, KS, in 2020 with the Arthropod-Borne Animal Diseases Research Unit (ABADRU). He transferred to the Foreign Arthropod-Borne Animal Diseases Research Unit (FABADRU) at the National Bio and Agro-defense Facility (NBAF) in 2020 before retiring in December 2024. He is a Marty Vanier and Bob Krause Biosecurity Research Institute Fellow and has served as a scientific consultant to the FAO/IAEA Joint Animal Health Division, as well as an Affiliate Faculty at Colorado State University, Texas A&M and Washington State University. He currently serves as Adjunct Professor at Kansas State University and an Extraordinary Lecturer at the University of Pretoria, South Africa. He has been a member of several professional societies including AAVLD since 1999 and continues to serve as the Membership Committee Chair for the World Society of Virology. Dr. Wilson's research focuses on the molecular biology, pathogenesis and diagnostics of arboviruses that cause important transboundary animal diseases (TADs) including bluetongue, Japanese encephalitis, Rift Valley fever and Vesicular Stomatitis. His work involves collaborative projects with national and international partners, including scientists from academia, government, and the bioscience industry in countries such as the US, Australia, Africa, and Japan. Recent collaborative partners include scientists from Kansas State University, the Canadian Food Inspection Agency, the Onderstepoort Veterinary Institute and the University of Pretoria in South Africa, as well as, in Kenya, the International Livestock Research Institute, the Kenya Agricultural Livestock Research Organization, the Kenya Department of Veterinary Services, and the Kenya Wildlife Services. Dr. Wilson's scientific activities have generated diagnostic tools and vaccines for the livestock industries as well as 10 book chapters and more than 150 peer-reviewed manuscripts.

Bio for Josh Daniels, DVM, PhD, DACVM (Bacteriology/Mycology)

Giles Scientific Excellence in Veterinary Diagnostic Microbiology Award 2025

Josh Daniels is a diagnostic microbiologist and has served as section head of the Bacteriology/Mycology section of The Colorado State University Veterinary Diagnostic Laboratory since 2017. Prior to his time at Colorado State, Josh was the head of Clinical Microbiology at The Ohio State University Veterinary Teaching Hospital from 2008-2017. Josh trained at The University of Wisconsin and Washington State University for his DVM and PhD respectively. He has authored or contributed to over 75 published manuscripts and book chapters.

Josh's passions are quality, clinical relevance of reported results, client education, and antimicrobial stewardship in the laboratory and on the clinic floor. As a teacher of future DVMs, his passion in the classroom is making Bacteriology less intimidating by writing songs about bacteria and getting students to sing along and come up with microbiology song lyrics of their own.

Karen Olsen is a Minnesota native and graduate of the University of Minnesota, where she earned her degree in Microbiology. She has devoted 38 years to the University of Minnesota Veterinary Diagnostic Laboratory, beginning as a technician in the bacteriology lab and working her way up to Section Manager. Over the years, she has focused on advancing diagnostic testing, supporting animal health, and mentoring the next generation of laboratory professionals. Outside of work, Karen enjoys traveling, birding, and spending time with her dog Archie.

Dr. Suzanna Storms, DVM, PhD, is a Clinical Microbiology Resident at the University of Illinois Veterinary Diagnostic Laboratory. Her professional focus includes diagnostic assay development and infectious disease surveillance in livestock populations. Dr. Storms has experience in virology and molecular diagnostics, with a special interest in influenza A virus and the development of improved tools for early and accurate detection of infectious diseases.

Hao Tong received his bachelor's degree in veterinary medicine from Huazhong Agricultural University in China in 2015. From 2015 to 2018, he worked at his family farm as a veterinarian. In 2018, he began pursuing an M.S. in Veterinary Preventive Medicine at the VDPAM department at Iowa State University. Afterwards, he returned to China, where he established and managed an on-site PCR laboratory to support disease prevention and eradication efforts on his family farm. In 2023, he returned to the United States to join Dr. Jianqiang(JQ) Zhang's lab as a Ph.D. student at Iowa State University.

His research interests include all swine diseases, particularly viral diseases. His PhD research mainly focuses on comparing the efficacy of multiple PRRSV vaccines and investigating the immunological and molecular factors contributing to the enhanced effectiveness observed

Mostafa Shelkamy is a PhD candidate in Veterinary Microbiology and Preventive Medicine at Iowa State University, working with the ISU poultry team. His research focuses on developing and improving molecular diagnostic tools for avian pathogens, with an emphasis on *Avibacterium paragallinarum*, the causative agent of infectious coryza. He has contributed to the discovery of non-pathogenic strains that may serve as live vaccines. Mostafa has authored several manuscripts and co-authored grants totaling nearly \$800,000. Mostafa is the recipient of multiple awards, including the 2024 AAAP Foundation Y.M. Saif Poultry Scholarship and the 2025 Lora and Russ Talbot Graduate Fellowship in Veterinary Medicine.

Ana Fiorella Castillo-Espinoza

Ana earned her DVM from the National University of San Marcos, Peru, and an MSc in Veterinary Preventive Medicine at Iowa State University (ISU), where she studied *Mycoplasma hyopneumoniae*–airway interactions using a three-dimensional (3D) tracheal model. She is currently a second-year PhD student in Veterinary Microbiology at ISU under Dr. Giménez-Lirola and Dr. Nelli, focusing on 3D cell culture models to study host–pathogen interactions of different *Mycoplasma* species and Influenza A Virus (IAV) subtypes, with potential applications in diagnostics. She also has over ten years of experience in veterinary diagnostics and remains active in research initiatives and academic activities in Peru.

Nneka Vivian Iduu is a doctoral researcher in the Biomedical Sciences Program at the College of Veterinary Medicine, Auburn University, with a focused interest in molecular diagnostics, antimicrobial resistance, and infectious diseases. Her work centers on developing and optimizing molecular diagnostic systems, using genomics and bioinformatic tools to improve pathogen detection and understanding, and to strengthen antimicrobial resistance surveillance and mitigation within a One Health framework. She has contributed to the development and validation of novel FRET-qPCR assays for the detection and differentiation of *Streptococcus equi* subspecies *equi* and *zooepidemicus*, along with assays targeting other emerging and persistent pathogens, through her work in the Molecular Diagnostics Laboratory with Dr. Chengming Wang. She remains committed to strengthening diagnostic capacity and supporting innovations that enhance both animal and human health outcomes.

Diba Yaghoubi is a graduate student in Dr. Viju Pillai's lab in the Department of Comparative Pathobiology at Purdue University College of Veterinary Medicine. I earned my bachelor's degree in cellular and molecular biology from Iran, followed by a master's degree in integrative physiopathology from École Pratique des Hautes Études – PSL Research University in Paris, France. I then completed a second master's degree in modelling and data analysis at Sorbonne University. Currently, as a graduate research assistant in the Pillai Lab, my research focuses on developing a rapid, field-deployable diagnostic test for the detection of mycoplasmosis in bison.

Calvin Ko is a mycoplasmologist and a veterinary pathologist dedicated to swine disease research and diagnostics. His specialties include bacterial culture and modern diagnostic tools application for fastidious Mollicutes, and he is now a Graduate Assistant Researcher and Anatomic Pathology Resident at Iowa State University. Currently, he is working with Dr. Rachel Derscheid and Dr. Pablo Pineyro with a focus on diagnostic method development, evaluation, and optimization in swine Mycoplasma, with training in diagnostic pathology.

Naomi Falconnier is an anatomic veterinary pathologist with a special interest in invertebrate pathology and molecular diagnostics. She studies infectious diseases of decapod crustaceans at the University of Florida as a graduate student. Although particularly fond of arthropods, her interests encompass all terrestrial and aquatic invertebrates.

Trevor Arunsiripate is a veterinarian in my third year of a combined anatomic pathology residency and PhD at Iowa State University's Veterinary Diagnostic Laboratory. All of my education has come from ISU. I am from undergrad, veterinary school, and now residency, and a graduate degree. My research interests are viral pathogens and their host interactions. And I LOVE the case flow at the diagnostic lab. Helping the submitting veterinarian and the farmer feels rewarding.

Margaret grew up in Seattle, Washington and earned her DVM at Cornell University. Prior to entering the field of pathology, she completed a small animal rotating internship in Seattle, and she is now a second-year anatomic pathology resident at the Texas A&M Veterinary Medical Diagnostic Laboratory.

Presentation: “Aberrant migration of *Angiostrongylus Cantonensis* in the central nervous system of a capybara.”

Naomi Falconnier is an anatomic veterinary pathologist with a special interest in invertebrate pathology and molecular diagnostics. She studies infectious diseases of decapod crustaceans at the University of Florida as a graduate student. Although particularly fond of arthropods, her interests encompass all terrestrial and aquatic invertebrates.

Dr. Agustin Rebollada-Merino is a veterinarian born and raised in an agricultural region of southwestern Spain, where the central role of animals in society inspired him to pursue a degree in Veterinary Medicine (DVM) and later a Ph.D. in Veterinary Sciences from the Complutense University of Madrid. His early work at the World Organization for Animal Health (WOAH) Reference Laboratory for Mammalian Tuberculosis, African Swine Fever, and African Horse Sickness (VISAVET) sparked a lasting interest in pathology and infectious disease investigation. He is currently a third-year, Senior Resident in Anatomic Pathology at the Animal Health Diagnostic Center, Cornell University. Agustin has participated in national and international research projects and contributed to scientific publications, book chapters, and conference presentations. His professional interests include bacterial pathogenesis, the development of diagnostic methods in pathology, and forensic pathology. He is committed to advancing these disciplines and to helping train the next generation of veterinarians.

I am a veterinarian born and raised in an agricultural region of southwestern Spain, where the central role of animals in society inspired me to pursue a degree in Veterinary Medicine (DVM) and later a Ph.D. in Veterinary Sciences from the Complutense University of Madrid. My early work at the World Organization for Animal Health (WOAH) Reference Laboratory for Mammalian Tuberculosis, African Swine Fever, and African Horse Sickness (VISAVET) sparked a passion for pathology and infectious disease investigation. I am currently a third-year, Senior Resident in Anatomic Pathology at the Animal Health Diagnostic Center, Cornell University. I have participated in national and international research projects and contributed to scientific publications, book chapters, and conference presentations. My professional interests include bacterial pathogenesis, the development of diagnostic methods in pathology, and forensic pathology. I am committed to advancing these disciplines and training the next generation of veterinarians.

Amy McKinney is a lab manager at the Athens Veterinary Diagnostic Laboratory at the University of Georgia, where she has worked for the past 17 years. She began her career as a bench technician in bacteriology, where she developed a passion for veterinary diagnostics and quality laboratory practices. Over the years, she has gained experience in multiple areas of the lab, including mycology, molecular biology, and most recently, next-generation sequencing (NGS). Amy is particularly interested in improving diagnostic workflows by streamlining sample processing, testing new methodologies, and implementing automation to reduce error rates and technician fatigue.

Her current work focuses on fungal whole-genome sequencing with the goal of identifying mutations associated with antifungal resistance. This project has involved evaluating DNA extraction methods and developing a reliable workflow for fungal pathogen identification in a veterinary diagnostic setting. Amy's career reflects a consistent commitment to innovation, quality, and advancing diagnostic capabilities in veterinary medicine.

Jingyi (Jenny) Li is a second-year anatomic pathology resident at the University of Illinois Urbana-Champaign. She earned her DVM from Tufts University Cummings School of Veterinary Medicine in 2024 and is originally from Shanghai, China. Her professional interests include diagnostic pathology, infectious and zoonotic diseases, and digital pathology. Jenny has been engaged in research since her undergraduate studies, contributing to projects on epilepsy using *Drosophila* model, the Common Loon microbiome, sex-based differences in SIV-infected, ART-suppressed macaques, and investigations of SARS-CoV-2 and monkeypox in rhesus macaques. She is currently completing her residency combined master's degree project on genome characterization and analysis of goat astrovirus. Looking ahead, she hopes to continue advancing her work in anatomic pathology and infectious diseases research.

Dr. Suzanna Storms, DVM, PhD, is a Clinical Microbiology Resident at the University of Illinois Veterinary Diagnostic Laboratory. Her professional focus includes diagnostic assay development and infectious disease surveillance in livestock populations. Dr. Storms has experience in virology and molecular diagnostics, with a special interest in influenza A virus and the development of improved tools for early and accurate detection of infectious diseases.

"Daniela Hidalgo-Montealegre, DVM, is a second-year master's student in Comparative Biomedical Sciences at The Ohio State University College of Veterinary Medicine. Her research focuses on antimicrobial resistance and the surveillance of multidrug-resistant pathogens, particularly Carbapenem-Resistant Enterobacterales. She currently serves as a Microbiology intern and member of the Infection Control Committee for the OSU Antimicrobial Stewardship Program at the Veterinary Medical Center. Under the mentorship of Dr. Dubraska Diaz-Campos and Dr. Thomas Wittum, she is developing expertise in clinical microbiology, infectious disease epidemiology, and antimicrobial stewardship. Daniela's professional interests include diagnostic stewardship, bacteriology, and the role of microbiology within One Health frameworks. Her work aims to strengthen surveillance systems and infection control strategies that address antimicrobial resistance across veterinary and human medicine."

Mostafa Shelkamy is a PhD candidate in Veterinary Microbiology and Preventive Medicine at Iowa State University, working with the ISU poultry team. His research focuses on developing and improving molecular diagnostic tools for avian pathogens, with an emphasis on *Avibacterium paragallinarum*, the causative agent of infectious coryza. He has contributed to the discovery of non-pathogenic strains that may serve as live vaccines. Mostafa has authored several manuscripts and co-authored grants totaling nearly \$800,000. Mostafa is the recipient of multiple awards, including the 2024 AAAP Foundation Y.M. Saif Poultry Scholarship and the 2025 Lora and Russ Talbot Graduate Fellowship in Veterinary Medicine.

MaRyka Smith is from the Center for Outcomes Research and Epidemiology within the College of Veterinary Medicine at Kansas State University. I am a PhD Candidate under Dr. Mike Sanderson. I completed my Doctor of Veterinary Medicine in May 2024 and hope to finish my PhD next spring. The core of my dissertation is epidemiology of Foot and Mouth Disease. My projects have specifically focused on reviewing biosecurity methods and assessing traceability of livestock in the event of an outbreak with computer modeling. I am exceedingly grateful for being selected for this award. I really enjoyed this conference and meeting so many future colleagues. I can't wait to attend again in the future.

Ana Fiorella Castillo-Espinoza

Ana earned her DVM from the National University of San Marcos, Peru, and an MSc in Veterinary Preventive Medicine at Iowa State University (ISU), where she studied *Mycoplasma hyopneumoniae*–airway interactions using a three-dimensional (3D) tracheal model. She is currently a second-year PhD student in Veterinary Microbiology at ISU under Dr. Giménez-Lirola and Dr. Nelli, focusing on 3D cell culture models to study host–pathogen interactions of different *Mycoplasma* species and Influenza A Virus (IAV) subtypes, with potential applications in diagnostics. She also has over ten years of experience in veterinary diagnostics and remains active in research initiatives and academic activities in Peru.

Maria Chaves is a PhD Candidate in Veterinary Microbiology at Iowa State University. She obtained her DVM in Brazil and moved to the United States in 2022 to pursue her dreams of higher education and improving her research skills. Her work focuses on the application of molecular diagnostic tools, specifically, Next Generation Sequencing Platforms for infectious agents' identification and characterization. She has been actively engaged in publishing her work in peer-reviewed journals and presenting her work across the country and internationally.

Calvin Ko is a mycoplasmologist and a veterinary pathologist dedicated to swine disease research and diagnostics. His specialties include bacterial culture and modern diagnostic tools application for fastidious Mollicutes, and he is now a Graduate Assistant Researcher and Anatomic Pathology Resident at Iowa State University. Currently, he is working with Dr. Rachel Derscheid and Dr. Pablo Pineyro with a focus on diagnostic method development, evaluation, and optimization in swine Mycoplasma, with training in diagnostic pathology.

Dr. Berenice Munguía-Ramírez obtained her DVM degree in México in 2019. She completed her MSc in Veterinary Preventive Medicine at Iowa State University in 2022, and currently, she is a PhD Candidate in Population Sciences in Animal Health at Iowa State University under the mentorship of Dr. Zimmerman and Dr. Giménez-Lirola.

Her research focuses on advancing molecular diagnostics for swine diseases and integrating diagnostics with swine epidemiology to improve disease detection and management strategies in the swine industry. Her work aims to enhance the accuracy of diagnostic methods, contributing to more effective disease control and prevention.

Pernille Knos Jorgensen (PJ) received her DVM from Copenhagen University in Denmark. She spent the first 5 years after graduation working in clinical practice in Norway and The United Kingdom. Following [TE1] this, she worked as a Veterinary Investigation Officer in Scotland for 6 years, performing necropsies on farm animals. PJ is especially fond of working with cattle. She is a second-year Anatomic Pathology Resident at the California Animal Health and Food Safety (CAHFS) Davis laboratory, University of California.

Konner Cool is a PhD student in the Department of Diagnostic Medicine/Pathobiology at Kansas State University's College of Veterinary Medicine. Under the mentorship of Dr. Juergen A. Richt, Konner has been working alongside peers and collaborators to establish ruminant animal models for SARS-CoV-2 and HPAIV H5N1 and to develop and validate SARS-CoV-2 specific diagnostic test systems for livestock and companion animals. As a fellow of the USDA-APHIS-NBAF Scientist Training Program (NSTP), Konner will join the Foreign Animal Disease Diagnostic Laboratory (FADDL) following graduation.

Steffi Coates was raised in Raleigh, North Carolina, surrounded by her parents, sister, three cats, two dogs, and two horses. She graduated from Davidson College in 2019 with a double major in Biology and German Studies.

Over the past two summers, Steffi has worked at the University of Kentucky Veterinary Diagnostic Laboratory with Dr. Laura Kennedy on retrospective analyses of Kentucky Thoroughbred racehorses, with a focus on humeral fractures and sesamoid bone pathology and imaging. She is deeply grateful for Dr. Kennedy's mentorship and support, which has been instrumental in shaping her research interests. Steffi is also thankful for the opportunity to present her work at the American Association of Veterinary Laboratory Diagnosticians (AAVLD) conference for the past two years, an experience that further strengthened her passion for pathology and equine research.

Sarah Harden Hiser, PhD/DVM and is a candidate at NC State's College of Veterinary Medicine, where she focuses on infectious disease dynamics and antimicrobial resistance. Her research centers on building AI-driven approaches that turn unstructured veterinary clinical data into tools for more proactive care, stewardship, and One Health surveillance. She has extensive experience in industry as a data scientist and previously worked as a principal data scientist and cloud architect at SAS and Deloitte. During her time leading the commercial field R&D team Sarah led conservation-focused AI projects on sea turtles, dolphins, sharks, and crop health. Sarah is passionate about advancing veterinary medicine through technology and equipping future veterinarians to integrate AI into their clinical practice.

Michelle Lund, DVM, is a candidate in the Class of 2029 at the Virginia-Maryland College of Veterinary Medicine, as well as a Registered Veterinary Technician. Before her time in veterinary medicine, Michelle was a corporate communications and strategic public relations expert, where she spent the majority of her first career working in aerospace and defense. Michelle holds a bachelor's degree in Communication Studies with a minor in Leadership Studies from Chapman University, a master's degree in Strategic Public Relations from The George Washington University, and completed the Veterinary Assisting certificate program and an associate of science degree in Veterinary Technology at the Community College of Baltimore County.

Cassie Paterson is a current first-year veterinary student at the University of Minnesota with interests in anatomic pathology and community medicine. She earned her Bachelor of Science in Biological Sciences at the University of Pittsburgh in 2022 and is originally from the Philadelphia area. She has worked as a necropsy technician at the University of Pennsylvania's New Bolton Center (PADLS), as a veterinary nurse in center city Philly, and has assisted in veterinary work at various zoos and emergency hospitals. In her spare time, Cassie enjoys spending time outdoors, reading novels, crocheting, baking gluten-free treats, and spending time with her wife and rat terrier mix named Nut!

Jus Tupik is a current third-year DVM student tracking public/corporate at the Virginia-Maryland College of Veterinary Medicine. She finished her PhD at Virginia Tech in 2024 on the innate immune response to Brucellosis and Lyme Disease. In Blacksburg, she enjoys working at the ViTALS AAVLD-accredited laboratory, hiking, and exploring new breweries. Jus plans to pursue careers in veterinary diagnostics to one day become a microbiology section head of a diagnostic lab.

Jiamao Ysng is a second-year student at Iowa State University CVM. Her hometown is Chongqing, a vibrant city of mountains, rivers and bridges, sitting in southeast of China. She has a white tabby cat and loves nature, food and dancing. She aims to be a veterinary diagnostic laboratory medicine researcher and educator specializing in next-generation sequencing (NGS), bridging clinical practice and advanced techniques to improve disease diagnosis and provide better service to the clients.